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ORIGINAL ARTICLE

An evaluation of some eco-friendly biopesticides against *Bemisia tabaci* on two greenhouse tomato varieties in Egypt

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Abstract

This study has two main approaches. First, it exploits the susceptibility of tomato cultivars as a prophylactic measure to detect auto resistance characters of the tested tomato varieties against *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). Secondly, it evaluates the efficacy of different bio-rational insecticides against *B. tabaci* under greenhouse conditions. The results exhibited a special significance in *B. tabaci* infestation suitability between the two tomato varieties with a high infestation significance found in the Shifa F1 hybrid tomato variety compared to the Savera F1 hybrid tomato variety in the first plantation period. Subsequently, in the second plantation period, there was a significant difference between the two tomato varieties. *Bemisia tabaci* showed a preference for the Shifa F1 hybrid over the Savera F1 hybrid tomato variety. These differences occurred during the 1st, 2nd, 4th, 6th, 7th, 8th, and 10th weeks. In the experimental trial for the efficacy of eco-friendly bio-rational insecticides, spinosad, azadirachtin, *Beauveria bassiana* and *Metarhizium anisopliae*, there were significant differences between the treated and untreated plants during the two plantation periods. A high efficacy of spinosad on the *B. tabaci* population was found. *Bemisia tabaci* infestation under all the applications was reduced from 50 to 94.61% for the two plantation periods. This obvious decrease in *B. tabaci* population increase attention to benefits of the different bio-rational insecticides.

Key words: *Bemisia tabaci*, bio-rational insecticides, greenhouse, susceptibility tomato

Introduction

Tomato, *Lycopersicon esculentum* Mill. is a very important vegetable crop throughout the world. Annually 107 million metric tons are produced, with fresh market tomatoes making up to 72% of the total (FAOSTAT 2002). Tomatoes are grown both under plastic covered greenhouses and in open fields. Tomatoes, wherever grown, are hosts for many kinds of insect pests: whiteflies, aphids, cabbage loppers, cutworms, tomato leaf miners, tomato fruit worms, tomato pinworm, mealy bugs and flea beetles (Abd El-Ghany 2011; Ibrahim *et al.* 2015). The whitefly *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae), a cosmopolitan insect pest, is one of the most destructive agricultural pests worldwide owing to its ability to feed on hundreds of plant species, many of which are important agricultural crops (Brown 2010; Dinsdale *et al.* 2010). Whiteflies are now globally distributed and are serious pests of cultivated crops in tropical and subtropical areas including Africa, Asia, Central America, South America, and the West Indies (Cock 1986; Chu *et al.* 2007). They are widespread throughout the Mediterranean region and have become increasingly important pests of cotton and vegetables in Egypt (De Barro *et al.* 2011).

The whitefly is a polyphagous insect pest on more than 600 different plant species (Oliveira *et al.* 2001; Bayhan *et al.* 2006; Stansly and Natwick 2010). It causes economic losses in vegetable, fiber, and ornamental crops due to both direct damage through phloem

feeding and injection of toxins and indirect damage to the host plant through its ability to transmit plant viruses (Pereira *et al.* 2004; Brown 2010). The direct damage elicited by *B. tabaci* has a vast impact on plant health and consequently yield. The indirect damage caused by the whitefly is even more destructive for agriculture (Lima *et al.* 2000). Indirect damage occurs through transmission of geminiviruses; whiteflies are vector of more than 300 plant viruses (Hogenhout *et al.* 2008). Moreover, an unfavorable side effect of whitefly infestation is the production of carbohydrate-rich honeydew excretions, which make the leaves sticky, impair photosynthesis and support the growth of sooty mold fungi on the plant leaf and fruit surface (Stansly and Natwick 2010). Whiteflies have a high level of resistance to chemical pesticides (Horowitz *et al.* 2005). Therefore, the present work aims to exploit the susceptibility of tomato cultivars as a prophylactic measure to its resistance characters against *B. tabaci* under greenhouse conditions. Moreover, the efficacy of different bio-rational insecticides against *B. tabaci* was evaluated.

Materials and Methods

Tomato varieties and plantation

The work was carried out in the greenhouse unit of the National Research Center (NRC) farm at El-Behira Governorate, District of Kom Hamada, about 160 km from Cairo. The farm covers 126 ha, of which about 7 ha make up a contiguous plot of greenhouses for vegetable crops. The average annual temperature of the area is around ±21.2°C, relative humidity (RH) around 57% and rainfall around 11.6 (±4.5). The soil is sandy, deep, and non-saline or slightly saline (EC values range between 0.2 and 0.5 $dS \cdot m^{-1}$). Field capacity and wilting point are low (8–9 and 2–3% of soil moisture content, respectively). Soil fertility has a low content of macro- and micronutrients and organic matter. Sprinkler and drip irrigation systems are used to irrigate field crops, vegetables and fruit trees. The principal source of irrigation water for the whole area is the Nubaria canal. The greenhouse is oriented north to south, and is covered by polyethylene plastic. The total area of the greenhouse is 480 m^2 (60 × 8 m). The soil was mixed with chick manure and tilled 3 times, once a week to increase fertility and reduce the viability of weed seeds and other possible pests. Drip irrigation pipes, 50 cm long, were installed between drippers to ensure even distribution of water. The irrigation system setup using waste water supplied by the organic fish farm. Plant growth was observed weekly during the whole plantation period to avoid any nutrient deficiencies. Chicken manure compost and organic fish farm water was sufficient enough to ensure proper plant growth. Hand picking of emerged weeds was carried out in the experimental greenhouse to prevent the competition between weeds and cultivated tomatoes.

Two different hybrid tomato varieties, Shifa and Savera, were bought from the Agriculture Research Center, Egypt (Shifa was imported from Thailand and Savera from China) and were sown on two different dates for the two different plantation periods (August 20 for the first plantation season and December 15, 2014 for the second plantation period in the horticulture nursery of the National Research Center). The tomato seeds were left for 30 days before transplanting under nursery conditions in order to obtain suitable germination and good plantlets before being transferred to the greenhouse. Tomato plantlets were transplanted on September 15, 2014 for the first plantation period and January 15, 2015 for the second plantation period. A strip plot design was used in the experimental greenhouse. It consisted of 3 replicates. Each replicate consisted of 6 double rows with a total of 12 rows for the block. The spaces between the double rows were 60–65 cm and the spaces between single rows ranged from 80 to 100 cm. The inter-row space was 50 cm as determined by the drippers in the pipeline. Spaces of 100–120 cm were left free between each replicate for experimental purposes. Each double row consisted of 40 transplants (20 plants per single row per block).

Field assay of *Bemisia tabaci* infestation suitability for the two tomato varieties

The purpose of this assay was to determine the difference in suitability of the studied tomato varieties for infestation by *B. tabaci* as a prophylactic measure. Sampling for *B. tabaci* was carried out after early detection using sticky traps (Yellow and Blue) for a period of 11 consecutive weeks, starting from early November 2014 to mid January 2015 for the first plantation period and from early March 2015 to mid May 2015 for the second plantation period, respectively. Twenty fully stretched new leaves for each variety were weekly inspected and observed in the greenhouse. The adults were counted for both tomato varieties in the greenhouse (Naranjo and Flint 1995).

Application of bio-rational insecticides

Four bio-rational insecticides were applied in this study namely, *B. bassiana* (Bio Power: Stanes Company), *Metarhizium anisopliae* (Bio Magic: Stanes Company), azadirachtin (Nimbecidine: Stanes Company), and spinosad (Tracer: Dow AgroSciences Company). The recommended dose given by the producer for field application was used and was assumed to be the median lethal concentration (LC_{50}). The median lethal

concentrations of bio-rational insecticides are: 5 ml · · l^{-1} · ha^{-1} for *B. bassiana;* 5 ml · l^{-1} · ha^{-1} for *M. anisopliae;* 5 ml · l^{-1} · ha^{-1} for azadirachtin and 0.3 ml · l^{-1} · ha^{-1} for spinosad. Additionally, higher and lower concentrations than the recommended doses (LC_{50}) were used. The higher concentration was the lethal dose (LD) and was prepared by doubling the LC_{50}. The lower concentration (SLD) was a double dilution to obtain the sub-lethal dose of LC_{25}. The efficacy of each insecticide at the three aforementioned concentrations was evaluated in the greenhouse trials. The three replicates were each divided into two sub replicates. Each concentration was applied on two double rows out of a total of six rows. A plastic sheet was used to separate the different treatments during application. All the sprayings were done at 7 day intervals. The applications of insecticides were done by using 5 l knapsack sprayers for each bio-rational insecticide. Evaluation of the tested bio-rational insecticides against *B. tabaci* was performed through random leaf sampling. Twenty infested leaves from different concentrations were observed in the greenhouse 6 days after insecticide application for the presence of *B. tabaci* adults on both tomato varieties for a period of 6 consecutive weeks, starting from early December 2014 to mid-January 2015 for the first plantation period and from early April 2015 to mid-May 2015 for the second plantation period.

Statistical analysis

An independent t-test was carried out to find the significant differences between the Shifa and Savera F1 hybrid tomato varieties in the suitability of *B. tabaci* based on the number of adults. The SPSS program Version 16 (SPSS-Inc. 2005) was used. The analysis of variance of the strip plot experiment for the results obtained from the efficacy of different bio-rational insecticides was carried out using the MSTAT-C Computer Software Program (MSTAT-C 1988). The least significant difference test was applied at 0.05 probability level to compare mean treatments.

Results and Discussion

Infestation suitability between tomato varieties under greenhouse conditions

Using a t-test a statistically significant difference in infestation suitability between two tomato F1 hybrid varieties, Shifa and Savera (2014–2015) under greenhouse conditions was found. Significantly high infestation was found in the Shifa F1 hybrid tomato variety compared to the Savera F1 hybrid tomato variety in the first season (Table 1). Significant differences were clearly seen during the 1st, 2nd, 3rd, 6th, 7th, 8th and 9th weeks. During the 4th, 5th, 10th and 11th weeks there were non-significant differences in infestation suitability between the two tomato varieties for the first plantation period. The greatest increase in infestation was recorded during the 7th and 8th weeks of the research and occurred during the 3rd and 4th weeks of December 2014 for both tomato varieties which had different levels of infestation (Fig. 1A). On the other hand, the lowest infestation was found in the two tomato varieties during the 1st, 2nd, 5th, 6th and 10th weeks of the

Table 1. *Bemisia tabaci* infestation suitability between Shifa and Savera tomato varieties under greenhouse conditions

No. weeks	1st plantation period			2nd plantation period		
	Shifa	Savera	p-value	Shifa	Savera	p-value
	no. adults (mean±SE)	no. adults (mean±SE)		no. adults (mean±SE)	no. adults (mean±SE)	
1	3.80±0.25	2.00±0.19	0***	3.65±0.23	2.40±0.37	0.006***
2	4.15±0.44	2.00±0.23	0***	5.25±0.49	3.50±0.22	0.002***
3	5.30±0.32	3.75±0.23	0***	5.60±0.62	4.45±0.26	0.95 ns
4	5.10±0.50	4.00±0.51	0.134 ns	7.65±0.52	5.60±0.27	0.001***
5	4.00±0.37	3.50±0.29	0.297 ns	6.75±0.76	5.75±0.47	0.272 ns
6	4.60±0.27	3.25±0.32	0.003***	7.80±0.51	6.20±0.39	0.017**
7	6.00±0.41	3.85±0.26	0***	7.35±0.50	4.75±0.38	0***
8	6.20±0.46	4.00±0.38	0.001***	8.20±0.55	5.15±0.47	0***
9	5.80±0.38	3.55±0.28	0***	8.05±0.85	5.70±0.44	0.20 ns
10	5.10±0.31	4.20±0.33	0.056*	6.50±0.38	4.75±0.30	0.001***
11	3.55±0.34	2.55±0.27	0.27 ns	5.15±0.48	4.45±0.38	0.262 ns

n = 20; *, **, ***, *t* is significant at the $p < 0.05$ level; ns – not significant

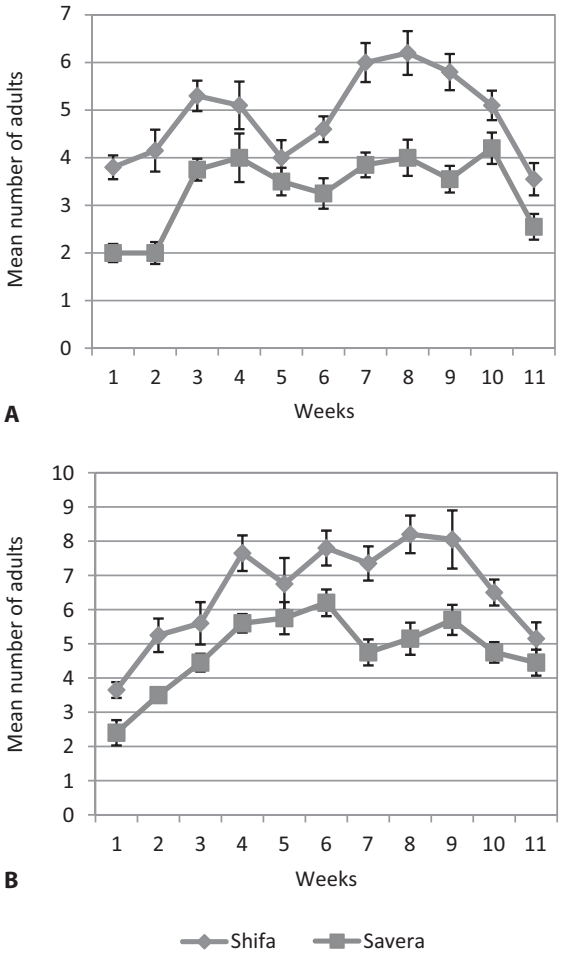

Fig. 1. Mean number of *Bemisia tabaci* adults between Shifa and Savera tomato varieties under greenhouse conditions: A – 1st plantation period and B – 2nd plantation period

experiment for the first plantation period. During the second plantation period, there was a significant difference between the two tomato varieties; *B. tabaci* showed a preference for the Shifa F1 hybrid over the Savera F1 hybrid tomato variety. The differences were seen during the 1st, 2nd, 4th, 6th, 7th, 8th, and 10th weeks. There were no significant differences between the two tomato F1 hybrid varieties during the 3rd, 5th, 9th and 11th weeks, respectively (Table 1). According to Figure 1B, the greatest infestation was recorded from March 17 to April 23, 2015. The lowest infestation levels were recorded during the 1st and 2nd weeks of March and the 1st and 2nd weeks of May 2015.

The data agree with Arnó *et al.* (2005) and Wen-Feng *et al.* (2009) who found that *B. tabaci* were more abundant in autumn than in spring and were very reduced during late winter. Moreover, Karut *et al.* (2012) reported that population densities of *B. tabaci* and infestation levels were higher in autumn than in spring (2008–2009 and 2009–2010). The differences in *B. tabaci* infestation levels between the Shifa F1 hybrid that have high infestation levels and the Savera F1 hybrid that have lower *B. tabaci* infestation levels could be due to the greater number of trichomes on the leaves and stems of the Savera variety than the number of trichomes on the leaves and stems of the Shifa variety. Cultivated tomato genotypes are covered with glandular and non-glandular types of trichomes on hypocotyls, stems, leaves, floral organs, and immature fruit. There is considerable diversity of trichome type, density and chemical composition within tomato species (Schilmiller *et al.* 2008). Numerous studies have shown that trichomes play an important role in the resistance of tomato species to insect infestation. Trichomes may contribute to resistance by impeding the ability of insects to feed, move, and survive on the plants (Kennedy 2003; Simmons and Gurr 2005). More importantly, resistance is also mediated by glandular trichome-borne metabolites that exert toxic effects on insect herbivores, or that physically entrap the insect upon rupture of the trichome gland. Differences in tomato infestation levels were reported by (JiRong and Ming 2011) because the hairy plant can repel aphids, *B. tabaci* and American leaf miners. Furthermore, Samarajeewa *et al.* (2005) found that the plants resistant to infestation with *B. tabaci* had more trichomes on their leaves and stems than the susceptible plants. Also, different compounds, such as acyl sugars, methyl ketones, terpenes, and alkaloids in tomato leaves, confer resistance to numerous insect pests of tomatoes, including aphids (*Macrosiphum euphorbiae* and *Myzus persicae*) and whitefly (*B. argentifolii*) (Rodriguez *et al.* 1993; Blauth *et al.* 1998).

Efficacy of bio-rational insecticides

In the present study, different bio-rational bio-pesticides were used to evaluate the efficacy of these bio-rational insecticides against *B. tabaci* adults under greenhouse conditions. The results from the 1st plantation period showed a significant difference between the treated and untreated (control) plants for Shifa and Savera F1 hybrid tomato varieties (Table 2 and Fig. 2A, B). These results could be easily calculated by the mean number of *B. tabaci* adults when treated with the different bio-rational insecticides for Shifa and Savera, respectively. With the different concentrations there were highly significant differences during the whole plantation period on Shifa and Savera F1 hybrid tomato varieties on the number of *B. tabaci* adults. The greater the concentration, the greater the efficacy on *B. tabaci* adult mortality gained. There were no significant differences between the LC_{50} and the higher concentrations of the applied bio-rational insecticides (Table 3).

In the 2nd plantation period there were statistically significant differences between the treated and untreated plants for both Shifa and Savera F1 hybrid tomato varieties with $p = 0$, during the six weeks of applications. These significant differences can easily be observed in Figure 2C, D for Shifa and Savera F1

Table 2. Efficacy interaction between different bio-rational insecticides against *Bemisia tabaci* adults

Variety		Shifa						Savera					
Investigation time / compound		week 1	week 2	week 3	week 4	week 5	week 6	week 1	week 2	week 3	week 4	week 5	week 6
1st plantation period	*Beauveria bassiana*	3.40	2.11	2.56	2.79	1.98	1.66	3.06	1.89	2.15	2.50	2.48	2.40
	Metarhizium anisopliae	3.18	1.93	2.51	2.60	1.76	1.41	2.93	1.78	2.03	2.18	1.54	1.40
	azadirachtin	4.28	2.23	2.71	3.08	1.96	1.55	3.56	1.90	2.49	2.40	2.39	2.39
	spinosad	3.80	2.21	2.85	2.83	1.95	1.56	3.68	1.84	2.06	2.25	1.50	1.46
F-value		***	0.30 ns	0.27 ns	0.18 ns	ns	ns	0.02**	ns	0.05	ns	0	0
LSD at 5%		0.13	0.08	0.08	0.10	0.08	0.08	0.12	0.08	0.08	0.10	0.08	0.07
2nd plantation period	*B. bassiana*	4.96	5.24	4.51	4.18	3.68	2.99	4.69	4.71	3.75	3.49	3.09	2.88
	M. anisopliae	5.59	5.08	4.65	4.10	3.60	3.20	5.20	4.78	3.65	3.48	3.08	2.89
	azadirachtin	3.88	2.93	3.25	2.98	2.29	1.65	3.11	2.28	2.46	1.88	1.30	1.30
	spinosad	2.93	2.78	2.45	2.21	1.71	1.24	2.59	1.96	1.83	1.68	1.28	1.05
F-value		0***	0***	0***	0***	0***	0***	0***	0***	0***	0***	0***	0***
LSD at 5%		0.08	0.07	0.08	0.05	0.06	0.08	0.09	0.08	0.07	0.08	0.08	0.08

n = 20; **, ***, *t* is significant at the $p < 0.05$ level; ns – not significant

hybrid tomato varieties, respectively. Furthermore, there were significant differences between the different concentrations of each bio-rational insecticide when compared with each other as shown in Tables 2 and 4. In spite of the significant differences between the different bio-rational insecticides in comparison to untreated plants, spinosad and azadirachtin were the strongest in their efficacy on adult *B. tabaci* during the two plantation periods on both Shifa and Savera F1 hybrid tomato varieties. *Metarhizium anisopliae* and *B. bassiana* showed highly significant efficacy during the first plantation period while during the second plantation period they tended to have a moderate efficacy on *B. tabaci* adults.

According to the Abbott formula (1925), spinosad reduced the greatest percent of *B. tabaci* adults at LC_{50} (from 83.97 to 94.61%) for both Shifa and Savera F1 hybrid tomato varieties under greenhouse conditions for the two plantation periods (Fig. 3). The next most effective was azadirachtin which gave an approximately 74.16 to 86.11% reduction, followed by *M. anisopliae* (52 to 89.94%) and finally *B. bassiana* with 50 to 87.65% efficacy on *B. tabaci*. There was a clear difference between the efficacies of the two entomopathogenic fungi during the two plantation periods. In the first plantation period *B. bassiana* and *M. anisopliae* achieved very satisfactory results on the *B. tabaci* adults. During the second plantation period *B. bassiana* and *M. anisopliae* caused almost 50 to 56.53% mortality of the *B. tabaci* adults.

Our results from the two plantation periods are similar to those of many authors who compared the efficacy of different microbial insecticides for integrated field management of the whitefly (Lacey *et al.* 2008; Lynn *et al.* 2010; Zhu and Kim 2011; Ghosh 2014; de Almeida Marques *et al.* 2014). Kim *et al.* (2011) studied the effect of different insecticides including spinosad on B and Q Bio-Types of *B. tabaci*. Their findings were comparable with what we found, showing the great efficacy of spinosad on *B. tabaci* populations. Moreover, Yin *et al.* (2011) investigated the efficacy of eight insecticides on *B. tabaci*. Their results showed that spinosad and Fipronil might be the best choices for the control of *B. tabaci* adults. Many researchers have investigated the efficacy of azadirachtin products against *B. tabaci* (Lynn *et al.* 2010; de Almeida Marques *et al.* 2014). De Almeida Marques *et al.* (2014) studied the effects of different plant extracts including neem for the management of *B. tabaci* Bio-Type B. A high efficacy of azadirachtin on *B. tabaci* infesting Solanaceae plants was found. However, Lynn *et al.* (2010) studied the effects of azadirachtin and neem based formulas on *B. tabaci*. They found a 78.2% reduction in the *B. tabaci* population which is in agreement with our results. Islam *et al.* (2011) also found the same significant effects of *B. bassiana* and azadirachtin on *B. tabaci*, namely, that there were no significant differences in mortality rates between the middle and sub-lethal doses used during their study.

Zhu and Kim (2011) reported different pathogenicity effects of different entomopathogenic fungi against *B. tabaci*. According to their results *B. bassiana* is very virulent and is considered to be one of the most virulent entomopathogenic fungi against *B. tabaci*. Lacey *et al.* (2008) studied the effect of the entomopathogenic fungi *B. bassiana* on the whitefly *B. tabaci*. They obtained

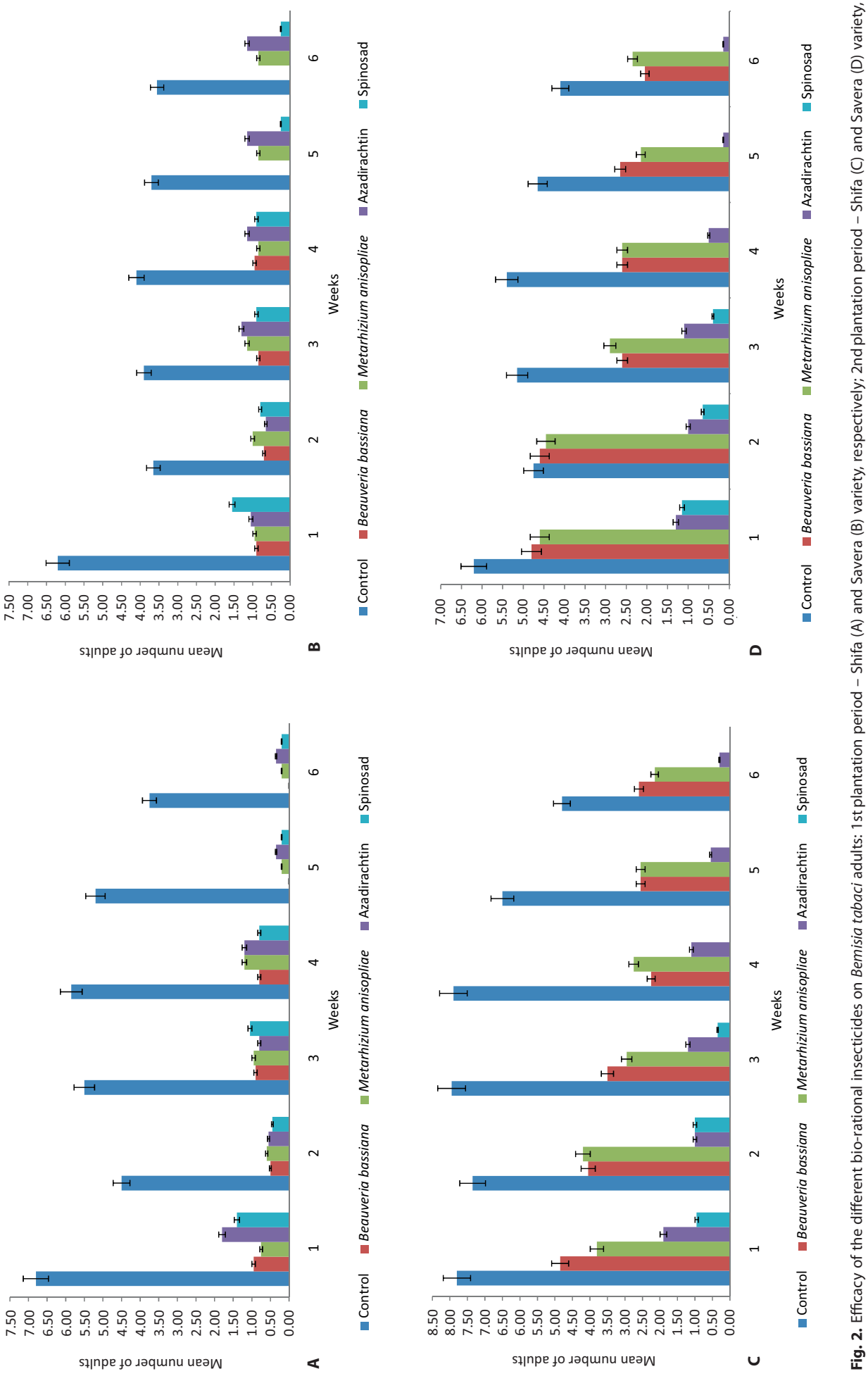

Fig. 2. Efficacy of the different bio-rational insecticides on *Bemisia tabaci* adults: 1st plantation period – Shifa (A) and Savera (B) variety, respectively; 2nd plantation period – Shifa (C) and Savera (D) variety, respectively

Table 3. Efficacy of different bio-rational insecticides on *Bemisia tabaci* adults for the 1st plantation period

Compounds	Variety	Shifa						Savera					
		week 1	week 2	week 3	week 4	week 5	week 6	week 1	week 2	week 3	week 4	week 5	week 6
	Concentration [ml · l^{-1} · ha^{-1}]	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults
Beauveria bassiana	control	6.80	4.50	5.50	5.85	5.20	3.75	6.20	3.65	3.90	4.10	3.70	3.55
	SLD	5.20	2.95	3.00	3.55	2.35	2.55	4.45	2.30	2.75	4.05	4.35	4.20
	MLD	0.75	0.60	0.95	1.20	0.20	0.20	0.95	1.00	1.15	0.85	0.85	0.85
	LD	0.85	0.40	0.80	0.55	0.15	0.15	0.65	0.60	0.80	1.00	1.00	1.00
Metarhizium anisopliae	control	6.80	4.50	5.50	5.85	5.20	3.75	6.20	3.65	3.90	4.10	3.70	3.55
	SLD	4.05	2.35	2.80	3.15	1.85	1.90	4.05	2.40	2.55	2.80	2.40	2.00
	MLD	0.95	0.50	0.90	0.80	0.00	0.00	0.90	0.70	0.85	0.95	0.00	0.00
	LD	0.90	0.35	0.85	0.60	0.00	0.00	0.55	0.35	0.80	0.85	0.05	0.05
Azadirachtin	control	6.80	4.50	5.50	5.85	5.20	3.75	6.20	3.65	3.90	4.10	3.70	3.55
	SLD	6.75	3.35	3.85	4.15	1.90	1.85	5.70	2.90	3.85	3.40	3.75	3.90
	MLD	1.80	0.55	0.80	1.20	0.35	0.35	1.05	0.65	1.30	1.15	1.15	1.15
	LD	1.75	0.50	0.70	1.10	0.40	0.25	1.30	0.40	0.90	0.95	0.95	0.95
Spinosad	control	6.80	4.50	5.50	5.85	5.20	3.75	6.20	3.65	3.90	4.10	3.70	3.55
	SLD	5.95	3.60	4.20	4.05	2.30	2.20	5.40	2.40	2.80	3.35	1.90	1.90
	MLD	1.40	0.45	1.05	0.80	0.20	0.20	1.55	0.80	0.90	0.90	0.25	0.25
	LD	1.05	0.30	0.65	0.60	0.10	0.10	1.55	0.50	0.65	0.65	0.15	0.15
F-value		0.12 ns	0.22 ns	0.07 ns	ns	ns	ns	ns	ns	0.27 ns	ns	0^{***}	0^{***}
LSD at 5%		0.24	0.14	0.16	0.18	0.16	0.16	0.23	0.14	0.14	0.19	0.16	0.15

n = 20; ***, *t* is significant at the $p < 0.05$ level, ns – not significant;
SLD – sub-lethal dose; MLD – median lethal dose; LD – lethal dose

Table 4. Efficacy of different bio-rational insecticides on *Bemisia tabaci* adults for the 2nd plantation period

Compounds	Variety	Shifa						Savera					
		week 1	week 2	week 3	week 4	week 5	week 6	week 1	week 2	week 3	week 4	week 5	week 6
	Concentration [ml · l^{-1} · ha^{-1}]	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults	mean no. adults
Beauveria bassiana	control	7.80	7.35	7.95	7.90	6.50	4.80	6.20	4.75	5.15	5.40	4.65	4.10
	SLD	4.60	5.50	3.90	3.65	3.30	2.95	4.80	5.45	4.05	3.30	3.20	3.05
	MLD	3.80	4.20	2.95	2.75	2.55	2.15	4.60	4.45	2.90	2.60	2.15	2.35
	LD	3.65	3.90	3.25	2.40	2.35	2.05	3.15	4.20	2.90	2.65	2.35	2.00
Metarhizium anisopliae	control	7.80	7.35	7.95	7.90	6.50	4.80	6.20	4.75	5.15	5.40	4.65	2.50
	SLD	5.55	5.40	3.95	3.75	3.10	3.15	5.25	5.70	3.75	3.45	2.85	2.45
	MLD	4.85	4.05	3.50	2.25	2.55	2.60	4.80	4.60	2.60	2.60	2.65	2.05
	LD	4.15	3.50	3.20	2.50	2.25	2.25	4.55	4.05	3.10	2.45	2.15	4.55
Azadirachtin	control	7.80	7.35	7.95	7.90	6.50	4.80	6.20	4.75	5.15	4.65	4.10	4.10
	SLD	4.15	2.35	2.70	1.70	1.55	1.20	3.50	2.50	2.50	1.75	0.95	0.95
	MLD	1.90	1.00	1.20	1.10	0.55	0.30	1.30	1.00	1.10	0.50	0.15	0.15
	LD	1.65	1.00	1.15	1.20	0.55	0.30	1.45	0.85	1.10	0.60	0.00	0.00
Spinosad	control	7.80	7.35	7.95	7.90	6.50	4.80	6.20	4.75	5.15	5.40	4.65	4.10
	SLD	2.20	2.00	1.10	0.95	0.35	0.15	2.45	1.95	1.30	1.30	0.45	0.10
	MLD	0.95	1.00	0.35	0.00	0.00	0.00	1.15	0.65	0.40	0.00	0.00	0.00
	LD	0.75	0.75	0.40	0.00	0.00	0.00	0.55	0.50	0.45	0.00	0.00	0.00
	F-value	0^{***}	0^{***}	0^{***}	0^{***}	0^{***}	0^{***}	0^{***}	0^{***}	0^{***}	0^{***}	0^{***}	0^{***}
	LSD at 5%	0.16	0.15	0.15	0.13	0.14	0.16	0.17	0.14	0.13	0.14	0.16	0.16

n = 20; ***, *t* is significant at the $p < 0.05$ level
SLD – sub-lethal dose; MLD – median lethal dose; LD – lethal dose

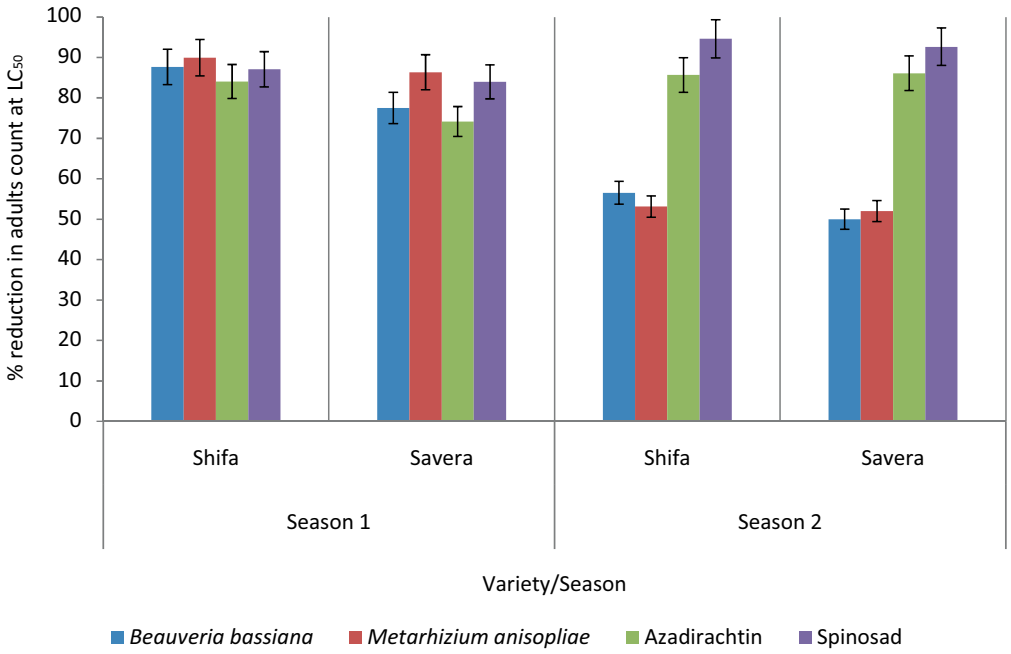

Fig. 3. Reduction percentage in *Bemisia tabaci* count at LC_{50} for different bio-rational insecticides

a reduction in the *B. tabaci* population of about 41 to 50% when compared with the untreated plants. Similar findings were obtained by Liu *et al.* (1999) and Ota *et al.* (1999) and were comparable to our results. Lacey *et al.* (2008) found that *M. anisopliae* reduced the *B. tabaci* population on eggplant by 30 to 90%. Batta (2003) obtained exactly the same results as we did. Santiago-Álvarez *et al.* (2006) also discovered the efficacy of *B. bassiana* on tomato and other host plants of *B. tabaci* to cause about 68 to 73% mortality of *B. tabaci*.

In conclusion, there were significant differences in infestation suitability between the studied varieties (Shifa and Savera) during the plantation periods. Significantly higher infestation was found in the Shifa F1 hybrid tomato variety than in the Savera F1 hybrid tomato variety. Therefore we recommend planting the Savera F1 hybrid tomato variety under greenhouse conditions during the tested periods. Concerning the utilization of bio-rational insecticides – previous research has indicated that when there are no control measures applied to tomato plants, there will be a 100% reduction in the produce. In our experimental trial for the evaluation of different bio-rational insecticides we found that spinosad, azadirachtin, *B. bassiana* and *M. anisopliae* had varied effects against *B. tabaci* on greenhouse tomatoes. The reduction in *B. tabaci* infestation under all the applications varied between 50 and 94.61% for the two plantation periods. Spinosad reduced the *B. tabaci* population by 83 to 94.61% and is ranked as the most effective bio-rational insecticide among the tested insecticides. It is followed by azadirachtin which reduced the *B. tabaci* population by 74 to 86.11%. *Metarhizium anisopliae* and *B. bassiana* gave a percent reduction of 52 to 89% and 50 to 87%, respectively. These different levels of efficacies on the target insect pest were translated into a 30–50% increase in total tomato produce when compared to the total harvested plants. This obvious increase merits paying attention to the cost benefits of the different bio-rational insecticides.

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References

Abbott W. 1925. A method of computing the effectiveness of an insecticide. Journal of Economic Entomology 18 (2): 265–267.

Abd El-Ghany N.M. 2011. Molecular evaluation of *Bacillus thuringiensis* isolates from the soil and production of transgenic tomato plants harboring Btgene for controlling lepidopterous insects in Egypt. Ph.D. thesis, Faculty of Science, Ain Shams University, Egypt, 270 pp.

Arnó J., Matas M., Martí M., Ariño J., Roig J., Gabarra R. 2005. Coexistence between *Trialeurodes vaporariorum* and *Bemisia tabaci* and impact of natural enemies in tomato crops under Mediterranean conditions. IOBC/WPRS Bulletin 28 (1): 1–4.

Batta Y. 2003. Production and testing of novel formulations of the entomopathogenic fungus *Metarhizium anisopliae* (Metschinkoff) Sorokin (Deuteromycotina: Hyphomycetes). Crop Protection 22 (2): 415–422.

Bayhan E., Ulusoy M.R., Brown J.K. 2006. Host range, distribution, and natural enemies of *Bemisia tabaci* 'B biotype' (Hemiptera: Aleyrodidae) in Turkey. Journal of Pest Science 79 (4): 233–240.

Blauth S.L., Churchill G.A., Mutschler M.A. 1998. Identification of quantitative trait loci associated with acylsugar accumulation using intraspecific populations of the wild tomato, *Lycopersicon pennellii*. Theoretical and Applied Genetics 96 (3): 458–467.

Brown J.K. 2010. Phylogenetic biology of the *Bemisia tabaci* sibling species group. p. 31–67. In: "*Bemisia:* Bionomics and Management of a Global Pest" (P.A. Stansly**,** S.E. Naranjo, eds.). Springer, 36 pp.

Chu D., Jiang T., Liu G., Jiang D., Tao Y., Fan Z., Zhou H., Bi Y. 2007. Biotype status and distribution of *Bemisia tabaci* (Hemiptera: Aleyrodidae) in Shandong province of China based on mitochondrial DNA markers. Environmental Entomology 36 (5): 1290–1295.

Cock M. 1986. *Bemisia tabaci,* a literature survey: on the cotton whitefly with an annotated bibliography. FAO/CAB, Ascot, United Kingdom, 121 pp.

de Almeida Marques M., Quintela E.D., Mascarin G.M., Fernandes P.M., Arthurs S.P. 2014. Management of *Bemisia tabaci* biotype B with botanical and mineral oils. Crop Protection 66: 127–132.

De Barro P.J., Liu S.S., Boykin L.M., Dinsdale A.B. 2011. *Bemisia tabaci*: a statement of species status. Annual Review of Entomology 56: 1–19.

Dinsdale A., Cook L., Riginos C., Buckley Y., De Barro P. 2010. Refined global analysis of *Bemisia tabaci* (Hemiptera: Sternorrhyncha: Aleyrodoidea: Aleyrodidae) mitochondrial cytochrome oxidase 1 to identify species level genetic boundaries. Annals of Entomology Society of America 103 (2): 196–208.

FAOSTAT. 2012. World to tomato production.

Ghosh S. 2014. Incidence of white fly (*Bemisia tabaci* Genn.) and their sustainable management by using biopesticides. p. 623–626. In: Proceedings of the 4th ISOFAR Scientific Conference at the 18th IFOAM Organic World Congress. Istanbul, Turkey, 13–14 October, 2014, 4 pp.

Hogenhout S.A., Ammar E.D., Whitfield A.E., Redinbaugh M.G. 2008. Insect vector interactions with persistently transmitted viruses. Annual Review of Phytopathology 46: 327–359.

Horowitz A.R., Kontsedalov S., Khasdan V., Ishaaya I. 2005. Biotypes B and Q of *Bemisia tabaci* and their relevance to neonicotinoid and pyriproxyfen resistance. Archives of Insect Biochemistry and Physiology 58 (4): 216–225.

Ibrahim S.S., Moharum F.A., Abd El-Ghany N.M. 2015. The cotton mealybug *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae) as a new insect pest on tomato plants in Egypt. Journal of Plant Protection Research 55 (1): 48–51.

Islam M.T., Omar D., Latif M., Morshed M.M. 2011. The integrated use of entomopathogenic fungus, *Beauveria bassiana* with botanical insecticide, neem against *Bemisia tabaci* on eggplant. African Journal of Microbiology Research 5 (21): 3409–3413.

JiRong Li W.H., Ming W.J. 2011. Breeding of 'Hangza No.401' – a new tomato cultivar with good quality and disease resistance. Journal of Anhui Agricultural University 38 (1): 110–117.

Karut K., Kazak C., Döker İ., Malik A.A.Y. 2012. Natural parasitism of *Bemisia tabaci* (Hemiptera: Aleyrodidae) by native Aphelinidae (Hymenoptera) parasitoids in tomato greenhouses in Mersin, Turkey. IOBC–WPRS Bulletin 80: 69–74.

Kennedy G.G. 2003. Tomato, pests, parasitoids, and predators: tritrophic interactions involving the genus Lycopersicon. Annual Review of Entomology 48: 51–72.

Kim S.I., Chae S.H., Youn H.S., Yeon S.H., Ahn Y.J. 2011. Contact and fumigant toxicity of plant essential oils and efficacy of spray formulations containing the oils against B-and Q-biotypes of *Bemisia tabaci*. Pest Management Science 67 (9): 1093–1099.

Lacey L.A., Wraight S.P., Kirk A.A. 2008. Entomopathogenic fungi for control of *Bemisia tabaci* biotype B: foreign exploration, research and implementation. p. 33–69. In: "Classical Biological Control of *Bemisia tabaci* in the United States – A Review of Interagency Research and Implementation" (J. Gould, K. Hoelmer, J. Goolsby, eds.). Springer, 343 pp.

Lima L., Návia D., Inglis P., De Oliveira M. 2000. Survey of *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae) biotypes in Brazil using RAPD markers. Genetics and Molecular Biology 23 (4): 781–785.

Liu T., Stansly P., Sparks J.A., Knowles T., Chu C. 1999. Application of Mycotrol and Naturalis-L (*Beauveria bassiana*) for management of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on vegetables, cotton and ornamentals in southern United States. Subtropical Plant Science: Journal of the Rio Grande Valley Horticultural Society 53: 44–48.

Lynn O.M., Song W.G., Shim J.K., Kim J.E., Lee K.Y. 2010. Effects on azadirachtin and neem-based formulations for the control of sweetpotato whitefly and root-knot nematode. Journal of the Korean Society for Applied Biological Chemistry 53 (5): 598–604.

MSTAT-C. 1988. MSTAT-C, a microcomputer program for the design, arrangement and analysis of agronomic research. Michigan State University, East Lansing.

Naranjo S.E., Flint H.M. 1995. Spatial distribution of adult *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton and development and validation of fixed-precision sampling plans for estimating population density. Environmental Entomology 24 (2): 261–270.

Oliveira M., Henneberry T., Anderson P. 2001. History, current status, and collaborative research projects for *Bemisia tabaci*. Crop Protection 20 (9): 709–723.

Ota M., Ozawa A., Kobayashi H. 1999. Efficacy of *Beauveria bassiana* preparation against whitefly on tomato. Annual Report of the Kanto-Tosan Plant Protection Society 46: 109–112.

Pereira M.F., Boiça J.A.L., Barbosa J.C. 2004. Distribuição espacial de *Bemisia tabaci* (Genn.) biótipo B (Hemiptera: Aleyrodidae) em feijoeiro (*Phaseolus vulgaris* L.). [Spacial distribution of *Bemisia tabaci* (Genn.) biotype B (Hemiptera: Aleyrodidae) in common bean (*Phaseolus vulgaris L.*)]. Neotropical Entomology 33 (4): 493–498.

Rodriguez A.E., Tingey W.M., Mutschler M.A. 1993. Acylsugars of *Lycopersicon pennellii* deter settling and feeding of the green peach aphid (Homoptera, Aphididae). Journal of Economic Entomology 86 (1): 34–39

Samarajeewa P., Meegahakumbura M., Rajapakse R., Gammulla C., Sumanasinghe V. 2005. Molecular and morphological identification of tomato yellow leaf curl virus (TYLCV) disease in tomato. Annuals of the SirLanka Department of Agriculture 7: 233–244.

Santiago-Álvarez C., Maranhão E.A., Maranhão E., Quesada-Moraga E. 2006. Host plant influences pathogenicity of *Beauveria bassiana* to *Bemisia tabaci* and its sporulation on cadavers. BioControl 51 (4): 519–532.

Schilmiller A.L., Last R.L., Pichersky E. 2008. Harnessing plant trichome biochemistry for the production of useful compounds. The Plant Journal 54 (4): 702–711.

Simmons A.T., Gurr G.M. 2005. Trichomes of *Lycopersicon* species and their hybrids: effects on pests and natural enemies. Agricultural and Forest Entomology 7 (4): 265–276.

SPSS-Inc. 2005. SPSS Base 16.0 for Windows User's Guide. SPSS Inc., Chicago IL.

Stansly P.A., Natwick E.T. 2010. Integrated systems for managing *Bemisia tabaci* in protected and open field agriculture. p. 467–497. In: "*Bemisia*: Bionomics and Management of a Global Pest (P.A. Stansly, S.E. Naranjo, eds.). Springer, 540 pp.

WenFeng Z., Peng T., Jinchang Ding Z.Y., Dong B.S., Guang G.J., ZengEn X. 2009. Temporal and spatial relationships among *Bemisia tabaci* and its natural enemies in tomato field. Journal of Journal Agricultural University 14 (4): 77–83.

Yin F., Feng X., Zhang D., Lin Q., Hu Z., Li Z., Chen H. 2011. Toxicities of 8 insecticides to *Bemisia tabaci* (Gennadius). Guangdong Agricultural Sciences 18: 025.

Zhu H., Kim J.J. 2011. Susceptibility of the tobacco whitefly, *Bemisia tabaci* (Hemiptera: Aleyrodidae) biotype Q to entomopathogenic fungi. Biocontrol Science of Technology 21 (12): 1471–1483.