



Journal of Plant Protection Research

eISSN 1899-007X

ORIGINAL ARTICLE

# Toxicity, antifeedant and repellent activities of five essential oils on adult *Cassida vittata* Vill. (Coleoptera: Chrysomelidae)

Achraf Charkaoui<sup>1</sup>\*<sup>o</sup>, Rachid Jbilou<sup>1</sup>, Houssam Annaz<sup>2,1</sup>, Kacem Rharrabe<sup>3,1</sup><sup>o</sup>

<sup>1</sup>Research Team Agricultural and Aquaculture Engineering, Polydisciplinary Faculty of Larache, Abdelmalek Essaadi University, Tetouan, Morocco

<sup>2</sup> Research Team Biotechnologies and Biomolecular Engineering, Faculty of Science and Technology Tangier, Abdelmalek Essaadi University, Tetouan, Morocco

<sup>3</sup>Research Laboratory Biology, Environment and Sustainable Development, Ecole Normale Superieure, Abdelmalek Essaadi University, Tetouan, Morocco

#### Vol. 64, No. 4: 323–335, 2024

DOI: 10.24425/jppr.2024.151820

Received: March 09, 2024 Accepted: May 08, 2024 Online publication: December 05, 2024

\*Corresponding address: acharkaoui@uae.ac.ma

Responsible Editor: Jacek Piszczek

#### Abstract

In Morocco, the sugar beet crop is severely harmed by the insect pest *Cassida vittata* Vill. which affects its yield quantity and quality. Chemical pesticides are considered the most common strategy to control this pest, and their use is extremely harmful to human health and the environment. In this context, the adults of C. vittata were exposed to five essential oils (EOs) obtained from: Artemisia herba alba Asso. (Asteraceae), Eucalyptus globulus Labill. (Myrtaceae), Mentha pulegium L. (Lamiaceae), Rosmarinus officinalis L. (Lamiaceae), and Shinus terebinthifolius Raddi. (Anacardiaciae). Their contact and fumigant activity was evaluated every 24 h for 3 days. Their repellent effect was tested by filter paper and sugar beet discs every 5 min for 30 min. Their antifeedant effect, via Relative Growth Rate (RGR), Relative Consumption Rate (RCR), Efficiency of Conversion of Ingested Food (ECI) and The Feeding Deterrence Index (FDI), was evaluated using three doses in each experiment. For the contact toxicity, M. pulegium, A. herba alba and R. officinalis showed the highest mortality rates with 100, 92 and 78%, respectively, after 24 h at 0.283  $\mu$ l · cm<sup>-2</sup>. For the fumigant toxicity, 100% mortality was observed at the highest concentration of M. pulegium after 24 h and from A. herba alba with 88 and 96 after 48 h and 72 h, respectively. Regarding the repellent effect by filter paper, the repellency of R. officinalis and A. herba alba was 82.92 and 57.85%, respectively. However, M. pulegium showed 63% of repellency after 5 min at 0.057  $\mu$ l · cm<sup>-2</sup>. In the antifeedant test, *M. pulegium* gave significant results in all nutritional indices. In conclusion, M. pulegium was the most effective in all tests used in this study. Our findings promote the use of these essential oils as efficient biocontrol compounds against the adults of C. vittata.

Keywords: bioinsecticide, Cassida vittata, essential oils, insect pest, toxicity

## Introduction

Sugar beet beetle *Cassida vittata* Vill. (Coleoptera, Chrysomelidae) is a widespread species commonly found in temperate climates in southern Europe as well as in northern Africa. However, in Morocco this insect causes extremely severe damage (Hmimina and Bendahou 2015). It is considered to be the principal insect pest of the sugar beet. Adults feed on the lower epidermis and inner tissues of the underside of sugar

beet leaves causing regular circular holes. They can also consume up to  $23.5 \text{ cm}^2$  per leaf (Fayed *et al.* 2014; Saleh *et al.* 2019) causing a strong loss of yield and decreasing the sugar content (Saleh *et al.* 2009; El-Dessouki *et al.* 2014; Hendawy and El-Fakharany 2017).

To control this insect in Morocco, a variety of conventional pesticides are used. Based on our investigations (unpublished data), the pesticides that are most



frequently used by farmers in the Larache region (northern Morocco) are Karate 5 EC against adults, and Dursban 4 EC against the larvae of C. vittata. The use of pesticides to eliminate this pest is extremely harmful to human health. Pesticides can lead to an increase in the risk of thyroid cancer (Norouzi et al. 2023), promote the occurrence and development of autoimmune diseases (Huang et al. 2023), negatively affect the blood-brain barrier (Cresto et al. 2023), raise the probability of infertility (Lahimer et al. 2023), increase the risk of cognitive decline (Sasaki et al. 2023), and can lead to cardiotoxicity (Marques et al. 2022). Furthermore, the toxicity of pesticides threatens aquatic environments (Eissa et al. 2023; Machate et al. 2023), leads to soil contamination (Liu et al. 2023) and could pose adverse risks towards honeybees (Li et al. 2023), sea turtles (Salvarani et al. 2023), birds (Cooke et al. 2023), fish (Wang et al. 2023) and freshwater (Nayak et al. 2023). Furthermore, 250,000 people die each year from pesticide poisoning (Pavela 2016).

These data highlight the urgent need to update the current pest control strategies, and find alternative, eco-friendly control methods to protect crops from herbivores. In this regard, natural products such as steroids, alkaloids, phenylpropanoids, phenolic compounds, terpenoids and nitrogenous compounds, as well as essential oils, emerge as great substitutes for synthetic pesticides with less to no harmful effects on human health and the environment (Gadban et al. 2020; Rharrabe et al. 2020; Kumar et al. 2021). More specifically, essential oils are becoming more and more popular as natural pest management solutions (Chang et al. 2022; Lima et al. 2023) since they are non-persistent in the environment and therefore less toxic to other insects and animals (Isman 2020). Among the advantages of essential oils, we may highlight the fact that they can act as repellents, attractants and antifeedants. They can also hinder insects' ability to recognize host plants and have acute toxicity (Isman 2006; Usseglio et al. 2023).

Based on recent studies, essential oils have been found to be effective against various species of Coleoptera, such as *Sitophilus zeamais* with essential oil from the leaves of *Schinus terebinthifolius* Raddi (Bernardi *et al.* 2024). *Eucalyptus globulus* L. (Myrtale: Myrtaceae) has been effective in the control of *Rhyzopertha dominica* (F.) (Siddique *et al.* 2022), *Callosobruchus maculatus* Fabricius has been affected by *Artemisia herba alba* Asso and *Rosmarinus officinalis L.* (Riffi *et al.* 2021; Baghouz *et al.* 2022). Furthermore, other species of insect pests such as *Thrips tabaci* Lindeman (Thysanoptera: Thripidae), *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae), and *Tuta absoluta* Meyer (Lepidoptera: Gelechiidae) were sensitive to the insecticidal effect of *Mentha pulegium* (Topuz 2023). The treatment of *C. vittata* by essential oil has not been documented and was one of the reasons why we started this work. We wanted to enrich the literature about the effect of essential oil on this insect, and to determine the toxicity, the repellent and the antifeedant effects of five essential oils obtained from *A. herba alba* Asso. (*Asteraceae*), *E. globulus* Labill. (*Myrtaceae*), *M. pulegium* L. (*Lamiaceae*), *R. officinalis* L. (*Lamiaceae*), and *S. terebinthifolius* Raddi. (*Anacardiaciae*) against adult beet beetles under laboratory conditions.

## **Materials and Methods**

#### **Essential oils**

*Eucalyptus globulus, Rosmarinus officinalis,* and *Mentha pulegium* essential oils were purchased from a farm in OUJDA region which specialized in the culture of aromatic and medicinal plants where essential oils are obtained by steam distillation. *Artemisia herba alba* was obtained from a farm in Bouachta (northern Morocco).

The essential oil of *Shinus terebinthifolius* was obtained by hydro distillation using a Clevenger-type apparatus. In brief, 200 g of fresh green grains of *S. terebinthifolius* was mixed with 500 ml of distilled water and subjected to 3 hours extraction after which the essential oil was separated from the hydrolats and mixed with a pinch of anhydrous sodium to eliminate any trace of water. All essential oils were stored in dark vials and kept in a refrigerator.

The chemical composition of the EOs had been previously assessed in our laboratory using GC-MS analysis. In summary, *Mentha pulegium* EO was dominated by the presence of pulegone (83.06%) (Moullamri *et al.* 2024). *E. globulus* and *R. officinalis*, as described by Ait-Ouazzou *et al.* (2011), were characterized by a slight increase in the major compound 1.8 cineole with a respective percentage of 84% and 46.10%. As for *S. terebinthilfolius* EO, α-pinene, p-cymene and limonene were the major compounds with percentages of 32.32, 16.74, and 11.26%, respectively. In *A. herba alba* EO, Camphor Thujone and B-thujone were identified as the major compounds with respective percentages of 32.04, 19.04, and 14.22%.

#### Insects

*Cassida vittata* adults were collected from infested leaves in sugar beet fields in Ksar el Kebir city (34°59'56"north, 5°54'10" west), between January and mid-June 2023. They were kept in a climate room with controlled parameters:  $25 \pm 1^{\circ}$ C,  $65 \pm 5^{\circ}$  relative humidity, and 16L:8D. They were placed with fresh sugar

beet leaves inside a box. For the experiment, the adults were chosen based on their age (more than 30 days). Adults were starved for 24 hours before being used in the experiment to encourage them to search for the food source.

#### **Contact toxicity**

The contact toxicity of the five essential oils was determined in an impregnated paper assay according to Obeng-Ofori *et al.* (1998) with some modifications. Three concentrations (0.6, 1.2, and 1.8%) prepared in acetone were used to screen the toxicity of the five essential oils, giving final concentrations of 0.094, 0.189, and 0.283  $\mu$ l · cm<sup>-2</sup>. One milliliter of each concentration was carefully distributed on filter paper (diameter 9 cm) with a micropipette. The solvent was allowed to evaporate for 3 min. The control received 1 ml of pure acetone. Filter paper was placed on the bottom of a Petri dish, then 10 adults of *C. vittata* were placed on the bottom of each Petri dish. Each treatment was replicated five times, and the mortality was recorded every 24 h for 3 days.

#### **Fumigant toxicity**

Fumigant toxicity was tested against adults of *C. vittata* according to Huang *et al.* (1997) with some modifications. The fumigant chamber consisted of 9 cm Petri dishes with a net volume of 120 ml. Two hundred and fifty  $\mu$ l of each concentration was distributed on the lid of a Petri dish and left to evaporate for 3 min. Ten adults were placed on the bottom of each dish and closed with the treated lid. Petri dishes were tightly sealed using parafilm. Mortality in adults was recorded every 24 h for 3 days. Three sets of acetonic concentrations (0.6, 1.2, and 1.8%) were prepared giving final concentrations of 12.5, 25, and 37.5  $\mu$ l · l<sup>-1</sup> of air, respectively. Five replicates were set for each concentration of essential oils and control.

#### **Repellent activity**

The repellent test was divided into two bioassays: the first one used filter paper, and the second one used leaf discs.

#### Filter paper bioassay

The repellent effect of the five essential oils against adult *C. vittata* was evaluated using the area preference method according to Talukder and Howse (1993) with some modifications. Filter paper circles (9 cm diameter) were cut into two halves. The first half received 300 µl of each concentration of essential oil while the second half received 300  $\mu$ l of acetone. Both halves were left for 3 min at room temperature to allow the evaporation of solvent. After evaporation, the two half-circles were glued together using duct tape and placed in a Petri dish (d = 9 cm). Then, 10 adults were placed in the center of each Petri dish which was then closed to avoid the escape of insects. Finally, the number of insects present in each half was counted after 5, 10, 15, 20, 25, and 30 min. Three concentrations (0.6, 1.2, and 1.8%) were used, giving final concentrations of 0.057, 0.113, and 0.170  $\mu$ l  $\cdot$  cm<sup>-2</sup>. Seven replicates were used for each treatment.

#### Leaf disc bioassay

The repellent effects of essential oils on adults were evaluated using the leaf disc test method (Cole 1994). Leaf discs, 1 cm in diameter, were treated; the first had 10  $\mu$ l of essential oil, and the second had 10  $\mu$ l of acetone. After 30 sec of evaporation, we placed them on two different sides of the Petri dish as illustrated in Figure 1. Thereafter, 10 adults were placed in the center of the dish and the number of insects in each zone was counted after 5, 10, 15, 20, 25, and 30 min, as described by Rharrabe *et al.* (2014). Three concentrations (0.6, 1.2, and 1.8%) were used, giving final concentrations of 0.076, 0.152, and 0.229  $\mu$ l  $\cdot$  cm<sup>-2</sup>. Seven replicates were used for each treatment.

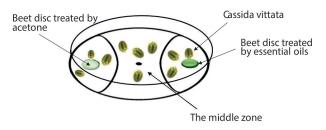


Fig. 1. Binary choices behavioral assay

For the two tests, the repulsion index was calculated according to Nerio Quintana *et al.* (2009) as follows:

$$PR = \frac{N_c - N_t}{N_c + N_t} \times 100$$

where:  $N_c$  – number of adults observed in the control area, and  $N_t$  – number of adults observed in the treated area. The mean repellency value of each extract was calculated and assigned to repellency classes from 0 to V (McDonald *et al.* 1970):

Class 0: The percentage of repulsion  $\leq 0.1\%$ : no repellency;

Class I:  $0.1\% < PR \le 20\%$ : very poor repellency; Class II:  $20.1\% < PR \le 40\%$ : moderate repellency;



Class III:  $40.1\% < PR \le 60\%$ : good repellency; Class IV:  $60.1\% < PR \le 80\%$ : very good repellency; Class V:  $80.1\% < PR \le 100\%$ : perfect repellency.

#### Antifeedant bioassay

The antifeedant activity was assessed using the nochoice test according to Schoonhoven (1982); discs (2-3 cm diameter) were cut from beet leaves. Discs and 10 adults were weighed separately. Next, 30 µl of each essential oil concentration were delicately added to the upper and lower sides of the discs. Leaf discs were left for 30 sec to allow the evaporation of acetone. Control discs received 30 µl of pure acetone. Finally, adults and treated discs were placed in a Petri dish and incubated at  $25 \pm 1^{\circ}$ C, 65% RH and 16: 8 h (L: D). After 3 days, both discs and insects were weighed again and the mass of insects and discs before and after the experiment were used to calculate the nutritional parameters (Manuwoto and Scriber 1985; Farrar et al. 1989). Three concentrations (0.6, 1.2, and 1.8%) were used for each essential oil giving final concentrations of 0.18, 0.36, and 0.54  $\mu$ l  $\cdot$  disc<sup>-1</sup>, and each treatment was conducted in seven replicates.

The nutritional parameters were calculated as follows:

Relative Growth Rate:  $BGB = \frac{A}{B}$ 

$$RGR = \frac{A - B}{B \times day},$$

where: A – weight of live insects (mg) on the third day/ number of live insects on the third day; B – original weight of insects (mg)/original number of insects.

Relative Consumption Rate (RCR):

$$RCR = \frac{D}{B \times day}$$

where: D – biomass ingested (mg)/number of live insects on the third day.

Efficacy of Conversion of Ingested food:

$$\mathrm{ECI} = \frac{\mathrm{RGR}}{\mathrm{RCR}} \times 100 \, .$$

The Feeding Deterrence Index (FDI):

$$FDI = \frac{C-T}{T} \times 100$$
 ,

where: C – consumption of control diet and T – consumption of treated diet.

#### **Statistical analysis**

Data were subjected to one-way analysis of variance (ANOVA) using SPSS version 25.0. Post-hoc testing was carried out using the Tukey's test. A significance level of 0.05 was used for all statistical tests.

## Results

#### **Contact toxicity**

The insecticidal activities of M. pulegium, A. herba alba, E. globulus, S. terebinthifolius and R. officinalis against C. vittata were examined by direct contact application (Fig. 2). Toxicity increased with increasing exposure periods. The highest effect was obtained with the essential oil from M. pulegium, followed by A. herba alba and R. officinalis. Regarding M. pulegium, we found that mortality started at the low concentration of 0.094  $\mu l \cdot cm^{-2}$  after 24 h with 54%. Then, when we raised the concentration to 0.189  $\mu$ l  $\cdot$  cm<sup>-2</sup>, we observed 86%, 88%, and 100% mortality after 24 h, 48 h, and 72 h, respectively. In addition, 100% mortality was recorded at the highest concentration used  $(0.283 \ \mu l \cdot cm^{-2})$  after 24 h. For the essential oil from A. herba alba, the mortality was between 52 and 58% at the lowest concentration (0.094  $\mu$ l  $\cdot$  cm<sup>-2</sup>) after 48 h and 72 h, respectively, and when we increased the concentration to 0.189  $\mu$ l · cm<sup>-2</sup>, the mortality increased to 70%, 76% and 90% after 24 h, 48 h and 76 h, respectively. For the EO from R. officinalis, the most effective activity was caused by using a concentration of 0.189  $\mu$ l · cm<sup>-2</sup>, showing 78, 94 and 100% mortality at 24, 48, and 72 h, respectively. Both EOs of *E. globulus* and S. terebinthifolius showed a low mortality percentage (<50%) against C. vittata, as shown in Figure 2

In the fumigation bioassay (Fig. 3) there were no significant differences between time and concentrations for *S. terebinthifolius* and *E. globulus* EOs.

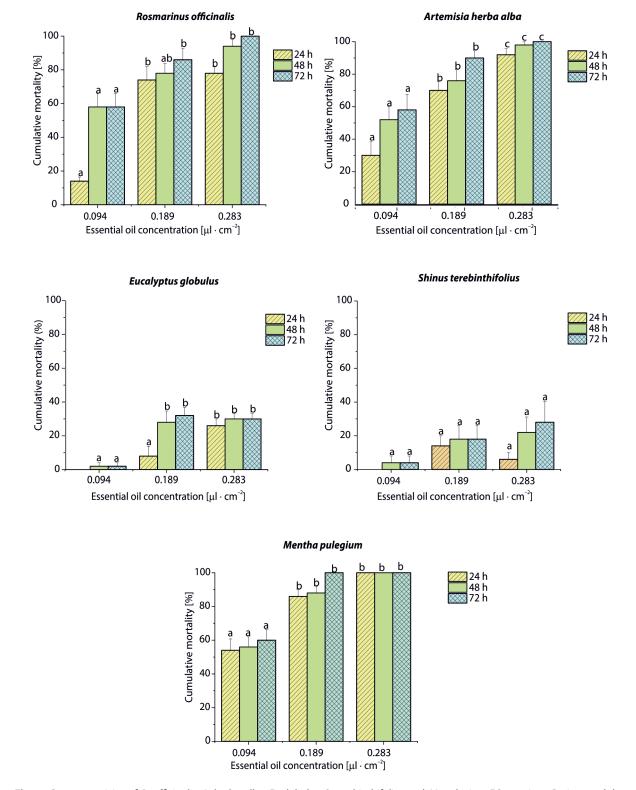
However, it was found that 64% mortality was due to *M. pulegium* at the concentration of 25  $\mu$ l · l<sup>-1</sup> of air during 24 h, 48 h and 72 h. In addition, 100% mortality was recorded at the highest concentration used  $(37.5 \ \mu l \cdot l^{-1} \text{ of air})$  after 24 h from the same essential oil. The toxicity of A. herba Alba was lower than *M. pulegium* with mortality rates of 6%, 88% and 96% at the highest concentration within 24 h, 48 h and 72 h, respectively. On the other hand, we found a significant mortality rate lower than 30% for R. officinalis. The toxicity of essential oils decreased in the following order: *M.* pulegium > *A*. herba alba > *R*. officinalis > *E*. globulus > S. terebinthifolius. EOs from M. pulegium and A. herba alba showed both significant contact and fumigant toxicity. R. officinalis showed the highest mortality rate only in the contact toxicity.

#### **Repellent activity**

In the filter paper test (Fig. 4), of the four EOs tested, *R. officinalis* exhibited the best levels of repellency at all the concentrations used with a percentage of repellency (PR) belonging to Class V of more than







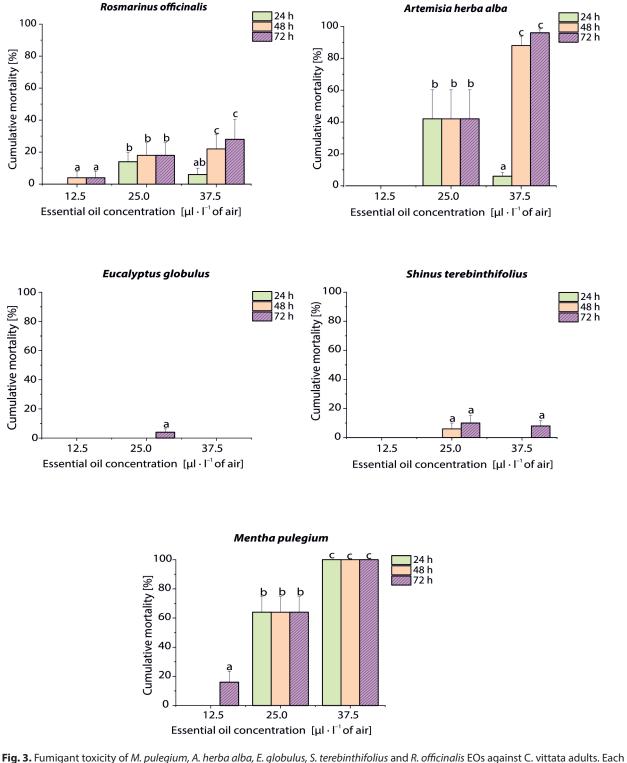
**Fig. 2.** Contact toxicity of *R. officinalis, A. herba alba, E. globulus, S. terebinthifolius* and *M. pulegium* EOs against *C. vittata* adults. Each point represents the mean ± SEM of five replicates. Different letters indicate statistical differences between the time and the concentrations among treatments while the same letter indicates no differences

80%. For *A. herba alba*, PR exceeded 60% at the lowest and the highest concentrations used, while 42.37% of repellency has been registered at the dose of 0.113  $\mu$ l · cm<sup>-2</sup> (hormetic effect), which demonstrated that this essential oil was a good repellent showing Class III repellency status. Furthermore, the repellency

of *M. pulegium* was more than 60% after 5 min at the lowest concentration (0.057  $\mu$ l · cm<sup>-2</sup>), and the repellency index decreased as we increased the concentration from 0.057  $\mu$ l · cm<sup>-2</sup> to 0.170  $\mu$ l · cm<sup>-2</sup>. On the other hand, both EOs of *E. globulus* and *S. terebinthifolius* showed a low repellent effect (<20%) against *C. vittata*.







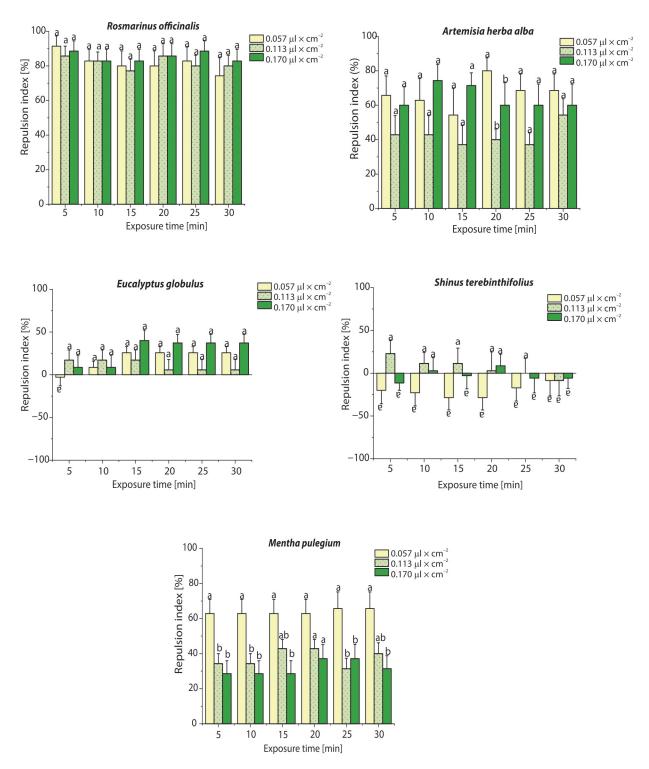
point represents the mean ± SEM of five replicates. Different letters indicate statistical differences between the time and the concentrations among treatments while the same letter indicates no differences

Regarding the repellent effect of the disc (Fig. 5), M. pulegium essential oil was very effective as a repellent against C. vittata adults with 65% repellency after 5 min exposure at 0.076  $\mu$ l  $\cdot$  cm<sup>-2</sup> doses, thereby being in Class IV. However, Classes II and III repellency status were observed at concentrations of 0.152  $\mu l \cdot cm^{\text{-2}}$ 

and 0.229  $\mu$ l · cm<sup>-2</sup>, respectively. On other hand, the repellency potential of A. herba alba corresponded to Class IV at the highest concentration used against the beetle. We noted an attractive effect (<20%) from the same EO at doses of 0.076  $\mu$ l  $\cdot$  cm<sup>-2</sup> and 0.152  $\mu$ l  $\cdot$  cm<sup>-2</sup>. Meanwhile, R. officinalis, S. terebinthifolius and E. glo-







**Fig. 4.** Repellent activity of *R. officinalis, A. herba alba, E. globulus, S. terebinthifolius* and *M. pulegium* EOs on C. vittata adults after different exposure times using a filter paper. Different letters indicate statistical differences between the time and the concentrations among treatments while the same letter indicates no differences

*bulus* were the least repellent EOs showing Classes I and II repellency status for most doses against the beetle *C. vittata* 

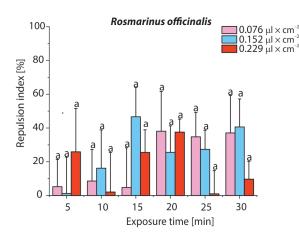
### **Antifeedant activity**

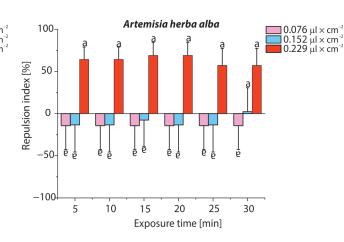
No significant differences in nutritional parameters were observed during the treatment with *E. globulus* 

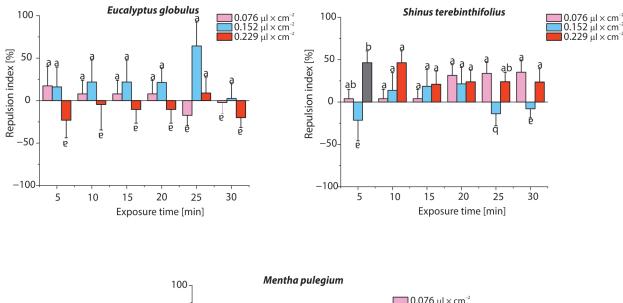
(Table 1). However, *S. terebinthifolius* had a significant reduction in RCR index compared to the control value at all concentrations used. Then, 56.4% of FDI occurred at 0.18  $\mu$ l × disc<sup>-1</sup> of concentration followed by 48.5 and 48% at 0.36  $\mu$ l · disc<sup>-1</sup> and 0.54  $\mu$ l · disc<sup>-1</sup> of concentration, respectively. For *R. officinalis*, we recorded a phagostimulant effect with –22.2% of the FDI at the two concentrations (0.18 and 0.36  $\mu$ l · disc<sup>-1</sup>)

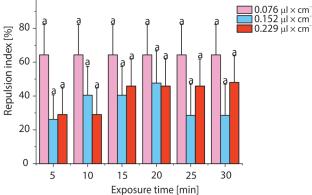


330









**Fig. 5.** Repellent activity of *R. officinalis, A. herba alba, E. globulus, S. terebinthifolius* and *M. pulegium* EOs on C. vittata adults after different exposure times using a disc of sugar beet. Each point represents the mean ± SEM of seven replicates. Different letters indicate statistical differences between the time and the concentrations among treatments while the same letter indicates no differences

followed by -29.2% at the highest concentration used. On the other hand, *M. pulegium* registered a significant effect on all nutritional parameters studied with a FDI value of 22.1% at the highest concentration used (0.54 µl · disc<sup>-1</sup>). Concerning *A. herba alba*, a significant difference was recorded only at the highest concentration used (0.54 µl · disc<sup>-1</sup>) for the RCR parameter.

## Discussion

The present study was the first attempt to test the toxicity by contact and fumigation as well as the repellent and antifeedant effects of five essential oils on *C. vittata* adults. Our results align with previous



**Table 1**. Antifeedant activity of EOs obtained from *R. officinalis, A. herba alba, S. terebinthifolius, M. pulegium* and *E. globulus*  $\pm$  SEM of seven replicates, at various concentrations on nutritional indices of *C.vittata* adults. Means within a column followed by the same letter are not significantly different (Tukey's HSD test, p < 0.05)

Plant Specie	Concentration $[\mu I \times disc^{-1}]$	RGR [mg/mg/day]	RCR [mg/mg/day]	ECI [%]	FDI [%]
Rosmarinus officinalis	0	-0.002 ± 0.002 a	0.571 ± 0.027 a	-0.2 ± 0.3 a	0.0 a
	0.18	$0.005 \pm 0.004$ a	0.721 ± 0.030 a	$0.7\pm0.5$ a	–22.2± 6.0 b
	0.36	$0.005 \pm 0.004$ a	$0.728 \pm 0.050 \text{ b}$	$0.6\pm0.5$ a	–22.1± 4.1 b
	0.54	$-0.001 \pm 0.005$ a	0.784 ± 0.046 b	$-0.2 \pm 0.7$ a	–29.2± 7.3 b
Artemisia herba alba	0	$-0.002 \pm 0.002$ a	0.571 ± 0.027 a	$-0.2\pm0.3$ a	0.000 a
	0.18	$-0.001 \pm 0.006$ a	$0.656 \pm 0.047$ a	$-0.4\pm0.9$ a	–19.6 ±6.4 a
	0.36	0.036± 0.021 a	0.723 ± 0.075 a	$4.4 \pm 2.7$ a	–22.0±8.3 a
	0.54	$-0.017 \pm 0.020$ a	0 .750 ± 0.055 b	$-3.0 \pm 2.7$ a	–38.4± 6.4 a
Schinus terebinthifolius	0	$-0.002 \pm 0.002$ a	0.571 ± 0.027 a	$-0.2\pm0.3$ a	0.0 a
	0.18	$0.015 \pm 0.009$ a	0.269 ± 0.017 b	$4.7\pm3.0a$	56.4 ± 3.1 b
	0.36	$-0.001 \pm 0.005$ a	0.269 ± 0.015 b	$0.7\pm1.9$ a	48.5 ± 4.2 b
	0.54	$0.006 \pm 0.009$ a	$0.288 \pm 0.019  b$	$0.8\pm3.8~a$	48.0 ± 2.2 b
Mentha pulegium	0	$-0.002 \pm 0.002$ a	0.571 ± 0.027 a	$-0.2\pm0.3$ a	0.0 ab
	0.18	$-0.005 \pm 0.002$ a	0.576 ± 0.029 a	$-0.8\pm0.3$ a	–6.0 ±4.0 b
	0.36	$-0.010 \pm 0.005$ ab	0.463 ± 0.01 b	$-2.4 \pm 1.1 \text{ ab}$	12.8 ± 3.7 ac
	0.54	$-0.060 \pm 0.040$ b	$0.349 \pm 0.05$ b	$-4.7 \pm 0.5$ b	22.1 ± 4.5 cd
Eucalyptus glo- bulus	0	$-0.002 \pm 0.002$ a	0.571 ± 0.027a	$-0.2\pm0.3$ a	0.0 a
	0.18	$-0.011 \pm 0.005$ a	$0.543 \pm 0.014a$	$-2.0\pm2.0$ a	$-3.6 \pm 2.8$ a
	0.36	$-0.006 \pm 0.002$ a	0.523 ± 0.021a	$-1.1 \pm 0.3$ a	$-5.4 \pm 2.6$ a
	0.54	$-0.040 \pm 0.046$ a	0.504 ± 0.047a	–13.1 ± 9.5 a	2.3 ± 9.1 a

findings by other researchers who have also tested the effects of the studied EOs on various Coleoptera species; *A. herba alba* demonstrated an important contact toxicity and fumigant toxicity against the adults of *Callosobruchus maculatus* Fab. (Aimad *et al.* 2022). Additionally, *M. pulegium* and *A. herba alba* showed the highest fumigant toxicity against *Tribolium confusum* (Abbad *et al.* 2014; Amoura *et al.* 2021), while *R. officinalis* exhibited contact and fumigant toxicity against adults of *Ephestia kuehniella* (Rekioua *et al.* 2022) and contact toxicity against *Sitophilus oryzae* (El-Bakry *et al.* 2019).

The results also showed that *M. pulegium* and *A. herba alba* EOs were the most repellent against *C. vittata* via the filter paper test and the disc test, respectively. According to the results found in the two tests, we noticed that the effectiveness of the essential oil was much stronger in the filter paper test than in the disc test. This result can be explained by the interaction of the odor given off by the beet plant, which may have weakened the effect of the essential oil used. This was shown by the insects' attraction to the beet disc, either by smell or color (Turlings and Ton 2006; Rharrabe *et al.* 2014; Arnold *et al.* 2015; Zhang *et al.* 2016; Webster and Card 2017). Moreover, *M. pule-gium* registered an indirect dose-response correlation

against *C.vittata*. We found that the repellency index of *M. pulegium* decreased as we increased the concentration. This may have been due to lateral inhibition within the antennal lobe, or to over-saturation of the receptor neurons of the insect (Skiri *et al.* 2004). The repellent effect of *M. pulegium* has been demonstrated on other insects, either as essential oils, against adult *Tribolium confusum* (Amoura *et al.* 2021) or as an extract against the two beetle pests *Tribolium castaneum and Lasioderma serricorne* (Salem *et al.* 2018) or as a powder of the aerial part against *Sitophilus oryzae* (Lougraimzi *et al.* 2018).

In the antifeedant test, we recorded a negative percentage of the FDI at the lowest concentration used by *R. officinalis*, which meant that the latter was a phagostimulant for *C. vittata*. In a different study, the extracts of *R. officinalis* had potent antifeedant activity against *Leptinotarsa decemlineata* Say (Sanchez-Vioque *et al.* 2015). Along the same lines, Kiran and Prakash (2015) saw that the essential oil of *R. officinalis* showed an antifeedant activity against *Sitophilus oryzae* L. (rice weevil) and *Oryzaephilus surinamensis* L. (sawtoothed grain beetle). For *M. pulegium*, we found a significant effect on all indices studied against *C. vittata*. Furthermore, an antifeedant effect was seen

by the same EO against *Tribolium castaneum* (Herbst) in another study by Heydarzade *et al.* (2019).

Among the EOs used, we found that *M. pulegium* was the most effective in all tests used against C. vittata adults. To explain the efficacy of this essential oil, numerous studies have shown that the most abundant compounds in *M. pulegium* are pulegone, menthone, and limonene (Herrera et al. 2014; Mejdoub et al. 2019; Candy et al. 2020; El Hassani 2020; Bachrouch et al. 2023). In keeping with that, numerous EOs rich in pulegone and menthone as the major compounds had insecticidal effects on other coleopters, such as T. confusum, Rhyzopertha dominica, Sitophilus zeamais and T. castaneum (Liu et al. 2011; Kasrati et al. 2015; Abbad et al. 2023). Also, they demonstrated both their contact and fumigant toxicity as well as a repellent effect against adults of Rhyzopertha dominica (Brahmi et al. 2016). Furthermore, the essential oil of Mentha piperita L., was also rich in menthone, menthol, and pulegone. These major compounds had a strong antifeedant effect against Callosobruchus maculatus (Saeidi and Mirfakhraie 2017).

The insecticidal property of *M. pulegium* EO is mainly attributed to major monoterpenoid compounds such as pulegone which are generally volatile and rather lipophilic and can penetrate rapidly into insects and interfere with their physiological functions (Bachrouch *et al.* 2015). Major compounds, such as pulegone and menthol can destroy insects by dysfunctioning the nervous system (Sánchez-Borzone *et al.* 2017; Jankowska *et al.* 2019a, b; Candy *et al.* 2020; Boulamtat *et al.* 2020), or through neuromuscular action (Ramdani *et al.* 2021). They can also affect cytochrome P450, which plays a key role in the detoxification system of insects (Rossi et *al.* 2012).

In the current literature there are no reports on the activity of EOs on *C. vittata*, even though the effectiveness of these essential oils on other insects has been extensively studied. It is necessary to conduct additional experiments to confirm the efficiency of the chosen EOs *on C. vittata* larvae stages and to test their effectiveness in sugar beet fields against the *C. vittata* beetle.

# Conclusions

The findings of the current study demonstrated that all the studied EOs had insecticidal activity in at least one test, especially *M. pulegium*, *A. herba alba* and *R. officinalis*. However, *M. pulegium* was found to be the most effective essential oil since it showed significant efficacy in all of the tests conducted. Using these products to reduce the damage caused by this insect, controlling it through their antifeedant and repellent effects, and reducing insect populations through their toxicity can be remarkably advantageous. However, further studies are required to assess their safety for humans and the environment and to develop practical large-scale applications under field conditions against *C.vittata*.

#### Acknowledgments

The authors are grateful to Zouhair Jendoubi for improving the English.

#### References

- Abbad A., Kasrati A., Jamali C.A., Zeroual S., M'hamed T.B., Spooner-Hart R., Leach D. 2014. Insecticidal properties and chemical composition of essential oils of some aromatic herbs from Morocco. Natural Product Research 28 (24): 2338–2341. DOI: https://doi.org/10.1080/1 4786419.2014.936015
- Abbad I., Soulaimani B., Abbad A. 2023. Chemical composition, insecticidal and allelopathic properties of essential oils obtained from wild and cultivated Moroccan Satureja calamintha (L.). Journal of Natural Pesticide Research 3. DOI: https://doi.org/10.1016/j.napere.2023.100 021
- Aimad A., Bourhia M., Hana H., Sanae R., Salamatullah A.M., Soufan W., Rihan H.Z., Ouahmane L., Youness E.A., Noureddine E., Mohamed F. 2022. Essential Oils from Artemisia herba alba Asso., Maticaria Recutita L., and Dittrichia Viscosa L. (Asteraceae): A Promising Source of Eco-Friendly Agents to Control Callosobruchus maculatus Fab. Warehouse Pest. Journal of Chemistry 2022:1–14. DOI: https:// doi.org/10.1155/2022/2373460
- Ait-Ouazzou A., Lorán S., Bakkali M., Laglaoui A., Rota C., Herrera A., Pagán R., Conchello P. 2011. Chemical composition and antimicrobial activity of essential oils of *Thymus* algeriensis, Eucalyptus globulus and Rosmarinus officinalis from Morocco. Journal of the Science of Food and Agriculture 91 (14): 2643–2651. DOI: https: //doi.org/10.1002/ jsfa.4505
- Amoura M., Benabdallah A., Kilani-Morakchi S., Messaoud C. 2021. Fumigant and repellent potentials of *Mentha pulegium* L. and *Citrus limon* L. (Burm) essential oils against *Tribolium confusum* Duval. (Coleoptera: Tenebrionidae). Journal of Entomological Research 45 (1): 73–80. DOI: https:// doi.org/10.5958/0974-4576.2021.00012.8
- Arnold S.E.J., Stevenson P.C., Belmain S.R. 2015. Responses to colour and host odour cues in three cereal pest species, in the context of ecology and control. Bulletin of Entomological Research 105 (4): 417–425. DOI: https://doi. org/10.1017/S0007485315000346
- Bachrouch O., Ferjani N., Haouel S., Ben Jemaa J.M. 2015. Major compounds and insecticidal activities of two Tunisian *Artemisia* essential oils toward two major coleopteran pests. Indus trial Crops and Products 65: 127–133. DOI: https:// doi.org/10.1016/j.indcrop.2014.12.007
- Bachrouch O., Zarroug Y., Bourgou S., Charradi K., Sriti J., Msaada K., Jallouli S., Chaibi K., Hamdi S.H., Abderraba M., Ben Jemâa J.M. 2023. Pennyroyal Essential Oil as a Green Pesticide for *Tribolium castaneum* (Herbst) Management and its Effects on Substrate Quality and Acetyl ch olinesterase Inhibition. Journal of the Mexican Chemical Society 67 (2): 152–162. DOI: http s://doi.org/10.29356/ jmcs.v67i2.1875
- Baghouz A., Bouchelta Y., Es-safi I., Bourhia M., Abdelfattah E.M., Alarfaj A.A., Hirad A.H., Nafidi H.-A., Guemmouh R. 2022. Identification of volatile compounds and insecticid al activity of essential oils from *Origanum com*-

pactum Benth. and Rosmarinus officinalis L. against Callosobruchus maculatus (Fab.). Journal of Chemistry 2022: 7840409. DOI: https://doi.org/1 0.1155/ 2022/ 7840409

- Bernardi J.L., Ferreira J.A., Puton B.M.S., Camargo S.D., Dal Magro J., Junges A., Cansian R.L., Steffens C., Zeni J., Paroul N. 2024. Potential agrochemical applications of *Schinus terebinthifolius* essential oil. Journal of Stored Products Research 105: 102260. DOI: https:// doi.org/10.1016/j. jspr.2024.102260
- Boulamtat R., Lhaloui S., Sabraoui A., El-Fakhouri K., Oubayoucef A., Mesfioui A., El-Bouhssini M. 2020. Antifeedant and larvicidal activities of *Mentha pulegium* on chickpea pod borer *Helicoverpa armigera* (Lepidoptera: Noctuidae). International Journal of Tropical Insect Science 40 (1): 151– 156. DOI: https://doi.org/10.1007/s42690-019-00064-z
- Brahmi F., Abdenour A., Bruno M., Silvia P., Alessandra P., Danilo F., Drifa Y.-G., Fahmi E.M., Khodir M., Mohamed C. 2016. Chemical composition and in vitro antimicrobial, insecticidal and antioxidant activities of the essential oils of *Mentha pulegium* L. and *Mentha rotundifolia* (L.) Huds growing in Algeria. Industrial Crops and Products 88: 96– 105. DOI: https://doi.org/10.1016/j.indcrop.2016.03.002
- Candy K., Akhoundi M., Andriantsoanirina V., Durand R., Bruel C., Izri A. 2020. Essential oils as a potential treatment option for pediculosis. Planta Medica 86 (9): 619–630. DOI: https://doi.org/10.1055/a-1161-9189
- Chang Y., Harmon P.F., Treadwell D.D., Carrillo D., Sarkhosh A., Brecht J.K. 2022. Biocontrol potential of essential oils in organic horticulture systems: from farm to fork. Frontiers in Nutrition 8. DOI: https://doi.org/10.3389/fnut.2021.805138
- Cole M.D. 1994. Key antifungal, antibacterial and anti-insect assays—a critical review. Biochemical Systematics and Ecology 22 (8): 837–856. DOI: https://doi.org/10.1016/0305-1978 (94)9008 9-2
- Cooke R., Whiteley P., Death C., Weston M.A., Carter N., Scammell K., Yokochi K., Nguyen H., White J.G. 2023. Silent killers? The widespread exposure of predatory nocturnal birds to antic oagulant rodenticides. Science of the Total Environment 904. DOI: doi.org/10.1016/j.scitoten v.2023.16 6293
- Cresto N., Forner-Piquer I., Baig A., Chatterjee M., Perroy J., Goracci J., Marchi N. 2023. Pesticides at brain borders: Impact on the blood-brain barrier, neuroinflammation, and neurological risk trajectories. Chemosphere 324. DOI: https://doi.org/10.1016/j.chemosphere.2023.138251
- Eissa F, Alsherbeny S., El-Sawi S., Slaný M., Lee S.S., Shaheen S.M., Jamil T.S. 2023. Remediation of pesticides contaminated water using biowastes-derived carbon rich biochar. Chemosphere 3 40. DOI: https://doi.org/10.1016/j. chemosphere.2023.139819
- El Hassani F.Z. 2020. Characterization, activities, and ethnobotanical uses of *Mentha* species in Morocco. Heliyon 6 (11). DOI: https://doi.org/10.1016/j.heliyon.2020.e05480
- El-Bakry A.M., Youssef H.F., Abdel-Aziz N.F., Sammour E.A. 2019. Insecticidal potential of Ag-loaded 4A-zeolite and its formulations with *Rosmarinus officinalis* essential oil against rice weevil (*Sitophilus oryzae*) and lesser grain borer (*Rhyzopertha dominica*). Journal of Plant Protection Research 59 (3): 324–333. DOI: https://doi.org/10.24425/ jppr.2019.129741
- El-Dessouki S.A., El-Awady S.M., El-Khawass K.A.M.H., Mesbah A.H., El-Dessouki W.A.A. 2014. Population fluctuation of some insect pests infesting sugar beet and the associated predatory insects at Kafr El-Sheikh Governorate. Annals of Agricultural Sciences 59 (1): 119–123. DOI: https://doi. org/10.1016/j.aoas.2014.06.016
- Farrar R.R., Barbour J.D., Kennedy G.G. 1989. Quantifying Food Consumption and Growth in Insect s. Annals of the Entomological Society of America 82 (5): 593–598. DOI: https://doi.org/10.1 093/aesa/82.5.593
- Fayed A.M., El-Magd B.M.A., Bazazo K.G.I., Mashaal R.E.F. 2014. Molecular and biochemical markers associated with

tolerance to *Cassida vittata* Vill (Coleoptera: Chrysomelidae) infestat ions in sugar beet. Egyptian Journal of Genetics And Cytology 43 (2): 393–406. DOI: 10.21608 / ejc.2014.9929

- Gadban L.C., Camiletti B.X., Bigatton E.D., Distéfano S.G., Lucini E.I. 2020. Combinations of *Tagetes filifolia* Lag. essential oil with chemical fungicides to control *Colletotrichum truncatum* and their effects on the biocontrol agent *Trichoderma harzianum*. Journal of Plant Protection Research 60 (1): 41–50. DOI: https://doi.org/10.24425/jppr. 2020.132202
- Hendawy A.S., El-Fakharany S.K.M. 2017. Parasitoids of the tortoise beetle, *Cassida vittata* vill. (coleoptera: Chrsomylidae) in sugar, fodder and table beet and effect of leaf phenols on parasi toid density. Egyptian Journal of Biological Pest Control 27 (2): 149–154.
- Herrera J.M., Zunino M.P., Massuh Y., Pizzollito R.P., Dambolena J.S., Gañan N.A., Zygadlo J.A. 2014. Fumigant toxicity of five essential oils rich in ketones against *Sitophilus zeamais* (Mots chulsky). AgriScientia 31 (1): 35–41. DOI: https:// doi.org/10.31047/1668.298x.v31.nl.9839
- Heydarzade A., Valizadegan O., Negahban M., Mehrkhou F. 2019. Efficacy of *Mentha spicata* and *Mentha pulegium* essential oil nanoformulation on mortality and physiology of *Tribolium castanetum* (Col.: Tenebrionidae). Journal of Crop Protection 8 (4): 501–520.
- Hmimina M., Bendahou S. 2015. The beet leaf beetle (*Cassida vittata* Wild, Col., Chrysomelidae) in Gharb: development cycle and control strategy. Revue Marocaine des Sciences Agronomiques et Vétérinaires 3 (3): 12–23. (in French)
- Huang R.-G., Li X.-B., Wang Y.-Y., Wu H., Li K.-D., Jin X., Du Y.-J., Wang H., Qian F.-Y., Li B.-Z. 2023. Endocrinedisrupting chemicals and autoimmune diseases. Environmental Research 231. DOI: https://doi.org/10.1016/j.envres.2023.116222
- Huang Y., Tan J.M.W.L., Kini R.M., Ho S.H. 1997. Toxic and antifeedant action of nutmeg oil against *Tribolium castaneum* (Herbst) and *Sitophilus zeamais* Motsch. Journal of Stored Products Rese arch 33 (4): 289–298. DOI: https:// doi.org/10.1016/S0022-474X(97)00009-X
- Isman M.B. 2006. Botanical insecticides, deterrents, and repellents in modern agriculture and an incr easingly regulated world. Annual Review of Entomology: 45–66. DOI: https:// doi.org/10.11 46/annurev.ento.51.110104.151146
- Isman M.B. 2020. Bioinsecticides based on plant essential oils: a short overview. Zeitschrift Fur Natur forschung Section C-a Journal of Biosciences 75 (7–8): 179–182. DOI: https:// doi.org/ 10.1515/znc-2020-0038
- Jankowska M., Lapied B., Jankowski W., Stankiewicz M. 2019a. The unusual action of essential oil component, menthol, in potentiating the effect of the carbamate insecticide, bendiocarb. Pestic ide Biochemistry and Physiology 158: 101–111. DOI: https://doi.org/10.1016/j.pestbp.2019.0 4.013
- Jankowska M., Wiśniewska J., Fałtynowicz Ł., Lapied B., Stankiewicz M. 2019b. Menthol Increases Bendiocarb Efficacy Through Activation of Octopamine Receptors and Protein Kinase A. Molecules 24 (20): 3775. DOI: https://doi. org/10.3390/molecules24203775
- Kasrati A., Jamali C.A., Bekkouche K., Spooner-Hart R., Leach D., Abbad A. 2015. Chemical Characterization and Insecticidal Properties of Essential Oils from Different Wild Populations of *Mentha suaveolens* subsp *timija* (Briq.) Harley from Morocco. Chemistry & Biodiversity 12 (5): 823–831. DOI: https://doi.org/10.1002/cbdv.201400236
- Kiran S., Prakash B. 2015. Toxicity and biochemical efficacy of chemically characterized *Rosmarinu s officinalis* essential oil against *Sitophilus oryzae* and *Oryzaephilus surinamensis*. Industrial Crops and Products 74: 817–823. DOI: https://doi. org/10.1016/j.indcrop.2015.05.073
- Kumar J., Ramlal A., Mallick D., Mishra V. 2021. An overview of some biopesticides and their importance in plant protection

for commercial acceptance. Plants-Basel 10 (6): 1185. DOI: httpps://doi.org/10.3390/plants10061185

- Lahimer M., Capelle S., Lefranc E., Cabry R., Montjean D., Bach V., Ajina M., Ali H.B., Benkhalifa M., Khorsi-Cauet H. 2023. Effect of pesticide exposure on human sperm characteristics, geno me integrity, and methylation profile analysis. Environmental Science and Pollution Research 30 (31): 77560-77567. DOI: https://doi.org/10.1007/s11356-023-27695-7
- Li W., Lv L., Wang Y., Zhu Y.-C. 2023. Mixture effects of thiamethoxam and seven pesticides with differrent modes of action on honey bees (Aplis mellifera). Scientific Reports 13 (1). DOI: htt ps://doi.org/10.1038/s41598-023-29837-w
- Lima A., Arruda F., Janeiro A., Medeiros J., Baptista J., Madruga J., Lima E. 2023. Biological activities of organic extracts and specialized metabolites from different parts of Cryptomeria japonica (Cupressaceae) - A critical review. Phytochemistry 206. DOI: https://doi.org/10.101 6/j.phytoch em.2022.113520
- Liu Z.L., Chu S.S., Jiang G.H. 2011. Toxicity of Schizonpeta multifida essential oil and its constituent compounds towards two grain storage insects. Journal of the Science of Food and Agriculture 91 (5): 905-909. DOI: https://doi.org/10.1002/ jsfa.4263
- Liu Y.-R., van der Heijden M.G.A., Riedo J., Sanz-Lazaro C., Eldridge D.J., Bastida F., Moreno-Ji ménez E., Zhou X.-Q., Hu H.-W., He J.-Z., Moreno J.L., Abades S., Alfaro F., Bamigboye A.R., Berdugo M., Blanco-Pastor J.L., de los Ríos A., Duran J., Grebenc T., Illán J.G., Makh alanyane T.P., Molina-Montenegro M.A., Nahberger T.U., Peñaloza-Bojacá G.F., Plaza C., Rey A., Rodríguez A., Siebe C., Teixido A.L., Casado-Coy N., Trivedi P., Torres-Díaz C., Verma J.P., Mukherjee A., Zeng X.-M., Wang L., Wang J., Zaady E., Zhou X., Huang Q., Tan W., Zhu Y.-G., Rillig M.C., Delgado-Baquerizo M. 2023. Soil contamination in nearby natural areas mirror s that in urban greenspaces worldwide. Nature Communications 14 (1). DOI: https:// doi.org/10.1038/s41467-023-37428-6
- Lougraimzi H., El Iraqui S., Bouaichi A., Gouit S., Achbani E.H., Fadli M. 2018. Insecticidal effect of essential oil and powder of Mentha pulegium L. leaves against Sitophilus oryzae (Linnaeus, 1763) and Tribolium castaneum (Herbst, 1797) (Coleoptera: Curculionidae, Tenebrionidae), the main pests of stored wheat in Morocco. Polish Journal of Entomology 87 (3): 263-278. DOI: https://doi.org/10.2478/pjen-2018 -0018
- Machate O., Schmeller D.S., Schulze T., Brack W. 2023. Review: mountain lakes as freshwater resources at risk from chemical pollution. Environmental Sciences Europe 35 (3). DOI: https://doi.org/10.1186/s12302-022-00710-3
- Manuwoto S., Scriber J.M. 1985. Differential effects of nitrogen fertilization of three corn genotypes on biomass and nitrogen utilization by the southern armyworm, Spodoptera eridania. Agricul ture, Ecosystems & Environment 14 (1): 25-40. DOI: https://doi.org/10.1016/0167-8 809(85) 90082-9
- Marques L.P., Joviano-Santos J.V., Souza D.S., Santos-Miranda A., Roman-Campos D. 2022. Cardiot oxicity of pyrethroids: molecular mechanisms and therapeutic options for acute and long-term toxicity. Biochemical Society Transactions 50 (6): 1737-1751. DOI: https://doi.org/10.1042/B ST20220593
- McDonald L.L., Guy R.H., Speirs R.D., 1970. Preliminary evaluation of new candidate materials as toxicants, repellents, and attractants against stored-product insects. Agricultural Research Service, US Department of Agriculture. Report number: 882. DOI: 10.22004/ag.Econ.3123 45
- Mejdoub K., Benomari F.Z., Djabou N., Dib M.E.A., Benyelles N.G., Costa J., Muselli A. 2019. Antifu ngal and insecticidal activities of essential oils of four Mentha species. Jundishapur Journal of Natural Pharmaceutical Products 14 (1). DOI: https://doi.org/10.5812/jjnpp.64165

- Moullamri M., Rharrabe K., Annaz H., Laglaoui A., Alibrando F., Cacciola F., Mondello L., Bouayad N., Zantar S., Bakkali M. 2024. Salvia officinalis, Lavandula angustifolia, and Mentha pulegium essential oils: insecticidal activities and feeding deterrence against Plodia interpunctella (Lepidoptera: Pyralidae). Journal of Essential Oil-Bearing Plants 27 (1): 16-33. DOI: https://doi.org/10.1080/097206 0X.2023.2301699
- Nayak S., Das S., Kumar R., Das I.I., Mohanty A.K., Sahoo L., Gokulakrishnan M., Sundaray J.K. 2023. Biochemical and histopathological alterations in freshwater fish, Labeo rohita (Hamilt on, 1822) upon chronic exposure to a commonly used hopper insecticide, triflumezopyrim. Chemosphere 337. DOI: https://doi.org/10.1016/j.chemosphere.2023.139128
- Nerio Quintana L.S., Olivero-Verbel J., Stashenko E. 2009. Repellent activity of essential oils from seven aromatic plants grown in Colombia against Sitophilus zeamais Motschulsky (Coleopter a). Journal of Stored Products Research 45: 212-214. DOI: https://doi.org/10.1016/j.jspr.200 9.01.002
- Norouzi F., Alizadeh I., Faraji M. 2023. Human exposure to pesticides and thyroid cancer: a worldwi de systematic review of the literatures. Thyroid Research 16 (1). DOI: https://doi. org/10.1186 /s130 44-023-00153-9
- Obeng-Ofori D., Reichmuth C.H., Bekele A.J., Hassanali A. 1998. Toxicity and protectant potential of camphor, a major component of essential oil of Ocimum kilimandscharicum, against four stored product beetles. International Journal of Pest Management 44 (4): 203-209. DOI: http s://doi.org /10.1 080/096708798228112
- Pavela R. 2016. History, presence and perspective of using plant extracts as commercial botanical insecticides and farm products for protection against insects - a review. Plant Protection Science 52 (4): 229-241. DOI: https://doi. org/10.17221/31/2016-PPS
- Ramdani C., El Fakhouri K., Sbaghi M., Bouharroud R., Boulamtat R., Aasfar A., Mesfioui A., El Bouhssini M. 2021. Chemical composition and insecticidal potential of six essential oils from Morocco against Dactylopius opuntiae (Cockerell) under field and laboratory conditions. Insects 12 (11): 1007. DOI: https://doi.org/10.3390/insects12111007
- Rekioua N., Boumendjel M., Taibi F., Samar M.F., Ben Jemaa J.M., Benaliouch F., Negro C., Nicoli F., De Bellis L., Boushih E., Haouel S. 2022. Insecticidal effect of Eucalyptus globulus and Rosmarinus officinalis essential oils on a stored food pest Ephestia kuehniella (Lepidoptera, Pyralidea). Cellular and Molecular Biology 68 (4): 144-157. DOI: https:// doi.org/10.14715/c mb/2022.68.4.18
- Rharrabe K., Jacquin-Joly E., Marion-Poll F. 2014. Electrophysiological and behavioral responses of Spodoptera littoralis caterpillars to attractive and repellent plant volatiles. Frontiers in Ecolog y and Evolution 2: 5. DOI: https://doi. org/10.3389/fevo.2014.00005
- Rharrabe K., Jbilou R., Bouayad N., Ajaha A., Aarab A. 2020. Harmaline ingestion effect on development, metabolites and midgut of the red flour beetle, Tribolium castaneum. Journal of Asia-Pacific Entomology 23 (1): 29-35. DOI: https://doi.org/10.1016/j.aspen.2019.10.013
- Riffi O., Fliou J., Elhourri M., El Idrissi M., Amechrouq A. 2021. Composition of the essential oil of the leaves of Artemisia herba alba Asso (Asteraceae) and its insecticidal activity on Callosobr uchus maculatus Fabricius (Coleoptera: Bruchidae). Journal of Microbiology, Biotechnology and Food Sciences 10 (6): e3293. DOI:10.15414/jmbfs.3293
- Rossi Y.E., Canavoso L., Palacios S.M. 2012. Molecular response of Musca domestica L. to Mintostachys verticillata essential oil, (4R) (+)-pulegone and menthone. Fitoterapia 83 (2): 33 6-342. DOI: https://doi.org/10.1016/j.fitote.2011.11.019
- Saeidi K., Mirfakhraie S. 2017. Chemical composition and insecticidal activity Mentha piperita L. essential oil against the cowpea seed beetle Callosobruchus maculatus F. (Coleoptera:

Bruchi da). Journal of Entomological and Acarological Research 49 (3). DOI: https://doi.org/10.4081 /jear.2 017.6769

- Saleh H.A., Khorchid A.M., El-Gably A.R. 2019. Efficiency of certain insecticides and their histologi cal effects against sugar beet beetle *Cassida vittata* (Coleoptera: Chrysomelidae) in sugar beet field. Egyptian Journal of Plant Protection Research Institute 2 (4): 751–758.
- Saleh M.M.E., Draz K.A.A., Mansour M.A., Hussein M.A., Zawrah M.F.M. 2009. Controlling the su gar beet beetle *Cassida vittata* with entomopathogenic nematodes. Journal of Pest Science 82 (3): 289–294. DOI: https://doi.org/10.1007/ s10340-009-0253-1
- Salem N., Sriti J., Bachrouch O., Msaada K., Khammassi S., Hammami M., Selmi S., Boushih E., Ouertani M., Hachani N., Abderraba M., Marzouk B., Limam F., Ben Jemaa J.M. 2018. Phenological stage effect on phenolic composition and repellent potential of *Mentha pulegiu m* against *Tribolium castaneum* and *Lasioderma serricorne*. Asian Pacific Journal of Tropical Biomedicine 8 (4): 207–216. DOI: https://doi.org/10.4103/2221-1691.231283
- Salvarani P.I., Vieira L.R., Rendón-von Osten J., Morgado F. 2023. Hawksbill Sea Turtle (*Eretmochel ys imbricata*) Blood and Eggs Organochlorine Pesticides Concentrations and Embryonic Deve lopment in a Nesting Area (Yucatan Peninsula, Mexico). Toxics 11 (1). DOI: https://doi.org/1 0.3390/toxics11010050
- Sánchez-Borzone M., Marin L., García D. 2017. Effects of insecticidal ketones present in mint plants on GA-BAA receptor from mammalian neurons. Pharmacognosy Magazine 13 (49): 114–117. DOI: https://doi.org/ 10.4103/0973-1296.197638
- Sanchez-Vioque R., Izquierdo-Melero M.E., Polissiou M., Astraka K., Tarantilis P.A., Herraiz-Penalver D., Martin-Bejerano M., Santana-Meridas O. 2015. Comparative chemistry and biol ogyical properties of the solid residues from hydrodistillation of Spanish populations of *Rosmarinus officinalis* L. Grasas Y Aceites 66 (2): e079. DOI: https://doi. org/10.3989/gya.1060142
- Sasaki N., Morse G., Jones L., Carpenter D.O. 2023. Effects of mixtures of polychlorinated biphenyls (PCBs) and three organochlorine pesticides on cognitive function differ between older Moh awks at Akwesasne and older adults in NHANES. Environmental Research 236. DOI: https:// doi. org/10.1016/j.envres.2023.116861
- Schoonhoven L.M. 1982. Biological aspects of antifeedants. Entomologia Experimentalis et Applicat a 31 (1): 57–69. DOI: https://doi.org/10.1111/j.1570758.1982.tb03119.x

- Skiri H.T., Galizia C.G., Mustaparta H. 2004. Representation of primary plant odorants in the antennal lobe of the moth *Heliothis virescens* using calcium imaging. Chemical Senses 29 (3): 253–2 67. DOI: https://doi.org/10.1093/chemse/ bjh026
- Siddique M.A., ul Hasan M., Sagheer M., Sahi S.T. 2022. Comparative toxic effects of *Eucalyptus globbulus* L. (Myrtales: Myrtaceae) and its green synthesized zinc oxide nanoparticles (ZnON Ps) against *Rhyzopertha dominica* (F.) (Coleoptera: Bostrichidae). International Journal of Tro pical Insect Science 42 (2): 1697–1706. DOI: https://doi.org /10.1007/ s42690-021-00691-5
- Talukder F.A., Howse P.E. 1993. Deterrent and insecticidal effects of extracts of pithraj, *Aphanamixis polystachya* (Meliaceae), against *Tribolium castaneum* in storage. Journal of Chemical Ecolog y 19 (11): 2463–2471. DOI: https://doi.org/10.1007/BF00980683
- Turlings T.C.J., Ton J. 2006. Exploiting scents of distress: the prospect of manipulating herbivore-induc ed plant odours to enhance the control of agricultural pests. Current Opinion in Plant Biology 9 (4): 421–427. DOI: https://doi. org/10.1016/j.pbi.2006.05.010
- Topuz E. 2023. Insecticidal activity of *Mentha pulegium* essential oil against *Thrips tabaci*, *Bemisia tabaci* and *tuta absoluta* adults. International Journal of Tropical Insect Science 43 (5): 1475–1483. DOI: https://doi.org/10.1007/s42690-023-01050-2
- Usseglio V.L., Dambolena J.S., Zunino M.P. 2023. Can essential oils be a natural alternative for the control of *Spodoptera frugiperda*? A review of toxicity methods and their modes of action. Plants 12 (1). DOI: https://doi.org/10.3390/ plants12010003
- Wang X., Chen F., Lu J., Wu M., Cheng J., Xu W., Li Z., Zhang Y. 2023. Developmental and cardiovascular toxicities of acetochlor and its chiral isomers in zebrafish embryos through oxidative stress. Science of the Total Environment 896. DOI: https://doi.org/10.1016/j.scitoten v.2023.165296
- Webster B., Card R.T. 2017. Use of habitat odour by host-seeking insects. Biological Reviews 92 (2): 1241–1249. DOI: https://doi.org/10.1111/brv.12281
- Zhang Z., Zhang M., Yan S., Wang G., Liu Y. 2016. A femalebiased odorant receptor from *Apolygus lucorum* (Meyer--Dur) tuned to some plant odors. International Journal of Molecular Sciences 17 (8): 1165. DOI: https://doi. org/10.3390/ijms17081165