

DOI 10.2478/v10181-012-0100-9

Original article

Participation of analogues of lysophosphatidic acid (LPA): oleoyl-sn-glycero-3-phosphate (L- α -LPA) and 1-oleoyl-2-*O*-methyl-*rac*-glycerophosphothionate (OMPT) in uterine smooth muscle contractility of the pregnant pigs

W. Markiewicz¹, K. Kamińska², M. Bogacki², T. Maślanka¹, J. Jaroszewski¹

¹ Department of Pharmacology and Toxicology, Faculty of Veterinary Medicine, University of Warmia nad Mazury, Oczapowskiego 13, 10-718 Olsztyn, Poland

² Institute of Animal Reproduction and Food Research, Polish Academy of Sciences, Tuwima 10, 10-747 Olsztyn, Poland

Abstract

Recent studies show that a representative of phospholipids, namely lysophosphatidic acid (LPA) and its receptors (LPA₁₋₃) play a significant role in the reproductive processes, i. a, in the modulation of the uterine contractility. The participation of LPA₃ in the reproductive processes has been revealed in mice and has not been studied in gilts. Therefore, in the present study we investigated the role/action of LPA and its receptors LPA₁, LPA₂ and LPA₃ on the contraction activity in the porcine uterus. The study was conducted on an experimental model in which the pig uterus consisted of the one whole uterine horn and a part of the second horn, both connected with the uterine corpus. Uterine strips consisting of the endometrium with the myometrium (ENDO/MYO) and myometrium (MYO) alone were collected on days 12-14 of the estrous cycle (control group; n = 5) or pregnancy (experimental group; n = 5). Two analogues of LPA at increasing doses were used: oleoyl-sn-glycero-3-phosphate (L- α -LPA, a selective agonist of LPA₁ and LPA₂ receptors; 10⁻⁷ M; 10⁻⁶ M and 10⁻⁵ M) and 1-oleoyl-2-*O*-methyl-*rac*-glycerophosphothionate (OMPT, a selective agonist of LPA₃ receptor; 68 nM; 136 nM and 680 nM). L- α -LPA caused an increase in the contraction tension, amplitude and frequency of ENDO/MYO from the uterine horn with the developing embryos. This effect was not observed in MYO in both groups examined. In the ENDO/MYO strips of the uterine horn with developing embryos, OMPT significantly increased the contraction tension at the highest dose (680 nM) and amplitude at all doses examined, while frequency of contractions was decreased at doses of 136 nM and 680 nM. In the MYO strips of the uterine horn with embryos a significant increase in the contraction tension and amplitude after the highest dose of OMPT was observed. The results obtained imply the important role of receptors LPA₁, LPA₂ and LPA₃ in the contraction activity of the porcine uterus during early pregnancy.

Key words: gilts, uterine smooth muscle, LPA, OMPT, embryos

Introduction

Spontaneous activity of the uterine myometrium, in terms of contraction tension, frequency and amplitude, is changing during the estrous cycle, mating or insemination, pregnancy and labour (Langendijk et al. 2002, 2005, Kurowicka et al. 2005). The contractile activity of the porcine myometrium is controlled by many factors including adrenergic, cholinergic and non-adrenergic non-cholinergic innervation (Łakomy 1987, Thilander et al. 1989, Taneike et al. 1991, Kaleczyc 1994, Majewski et al. 1995), as well as endocrine, paracrine and autocrine factors (Kitazawa et al. 1997, 2003, Cao et al. 2002, Sukjumlong et al. 2009, Franczak and Bogacki 2009). The most important substances regulating porcine uterine contractions include: noradrenaline (NA), serotonin (Kitazawa et al. 2001a), acetylcholine (Ach) (Kitazawa et al. 2003), oxytocin (Carnahan et al. 1999, Kitazawa et al. 2001b), prostanoids (prostaglandin $F_{2\alpha}$, E_1 and E_2) (Kucharski et al. 2007, Jana et al. 2010), thromboxane A_2 , histamine (Kitazawa et al. 1997), neuropeptide Y (Markiewicz et al. 2003) and endothelin (Isaka et al. 2000). Recently it has been documented that besides the above-mentioned substances, a representative of phospholipids, such as lysophosphatidic acid (1-acyl-*sn*-glycerol-3-phosphate- LPA) and its receptor (LPA_3) may play a significant role in reproductive processes including modulation of the uterine contractility (Tigyi et al. 1992, Tokumura et al. 1999, Budnik and Mukhopadhyay 2002). LPA is a multifunctional lipid messenger and is the simplest of all glycerophospholipids (Moolenaar 1995). Lysophospholipase D is the enzyme responsible for LPA production (Xie and Meier 2004). This bioactive lipid mediator acts via G-protein-coupled receptors evoking multiple effects including Ca^{2+} mobilization, chemotaxis and cell proliferation (Tigyi et al. 1994, Liliom et al. 1996a, 1996b). LPA has been detected in different types of fluids including serum, plasma, saliva and follicular fluid (Tokumura et al. 1986, 1999, Tigyi and Miledi 1992). Recent identification and genetic manipulations of G-protein-coupled receptors specific to LPA (LPA_{1-3}) have provided mechanistic and functional insights into their diverse roles in biological processes (An et al. 1998, Bandoh et al. 1999, Contos et al. 2000a, 2002, Noguchi et al. 2003, Gardell et al. 2006). For example, targeted disruption of LPA_1 and LPA_2 has demonstrated their physiological and pathological functions in the neural development, neuropathic pain and diarrhea (Contos et al. 2000b, Inoue et al. 2004, Li et al. 2005). LPA_3 receptor is involved in the contractility of the myometrium in mice what determines evenly embryo spacing along the uterine horns (Hama et al. 2007). The increased expression of LPA_3

in the porcine endometrium during the periimplantation period indicates that it plays an important role during this stage of pregnancy (Ye et al. 2005, Kaminska et al. 2008). In the present study we investigated whether LPA and its analogues participate in the uterine contraction in gilts in early pregnancy. Considering the fact that in the porcine uterus the transcripts for LPA_1 and LPA_2 were also identified (Seo et al. 2008), we studied whether these isoforms may participate in action of LPA on the porcine uterus contraction activity.

Materials and Methods

Prepubertal crossbred gilts ($n=10$) with an average body weight of 106 ± 4.8 kg and approximately 7 months of age were used. The gilts were subjected to surgical procedure under general anesthesia. The animals were premedicated with azaperone (Stresnil, Janssen Animal Health; 2 mg/kg i.m.), ketamine (VetaKetam, Vet-Agro, Poland; 10-15 mg/kg i.m.) and anaesthetized with thiopental (Sandoz, GMBH, Austria 20-30 mg/kg i.v.) In five gilts (experimental group) the uterine horns were presented by a midventral opening of the caudal part of the abdomen as described previously (Wasielak et al. 2008). Briefly, one horn of each gilt was cut transversely and the ends were closed by a suture. This way an experimental model was created in which the pig uterus consisted of the one whole uterine horn and a part of the second horn, both connected with the uterine corpus. The remaining part of the second horn, connected with the contiguous ovary, was surgically detached from the uterine corpus. After 10 days of recovery from surgery, gilts were treated hormonally by an intramuscular injection of 750 I.U. PMSG (Folligon, Intervet, Poland) and 500 I.U. of hCG (Chorulon, Intervet) given 72 h later. Subsequently, gilts from the experimental group ($n=5$) were inseminated 24 h after the hCG treatment. The insemination was repeated twice at 12 h intervals. The remaining gilts ($n=5$; control group) were subjected to surgical procedure but not inseminated. At the Days 12-14 of pregnancy (experimental group) or estrous cycle (control group) gilts were slaughtered. To confirm pregnancy, the uterine horns were flushed with 10 ml PBS (pH=7.4) to determine the presence of embryos in uterine flushings (Wasielak et al. 2008). Fragments of the uterine horns, collected from the middle part of the horns, were transferred to ice and transported to the laboratory and immediately processed for examination of the contractile activity. The contractile activity was examined according to the method described previously (Jana et al. 2010). Briefly, two kinds of the uter-

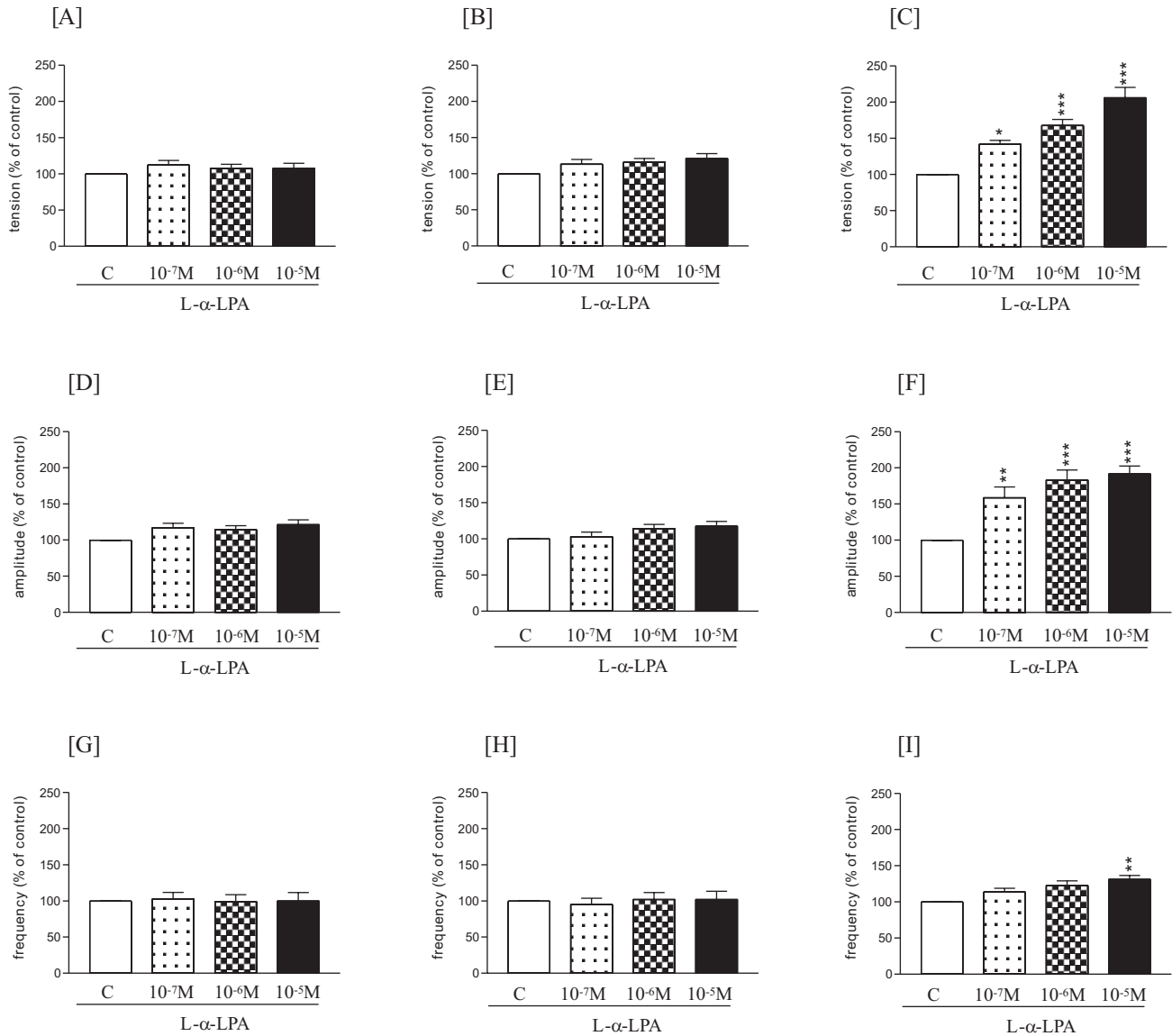


Fig. 1. Influence of oleoyl-sn-glycero-3-phosphate (L- α -LPA) on the tension ([A],[B],[C]) amplitude ([D],[E],[F]) and frequency ([G],[H],[I]) of the contraction of the porcine endometrium/myometrium strips of the control group ([A],[D],[G]), uterine horn without embryos ([B],[E],[H]) and with the embryos ([C],[F],[I]). Uterine strips were collected on days 12-14 of the estrous cycle (control group) or pregnancy (groups with embryos and without embryos). Values (mean \pm SEM; n = 5 in each group) are expressed as a percentage of changes in the contractile activity before the treatment. * P < 0.05, ** P < 0.01, *** P < 0.001 as compared to the contractile activity before the treatment (C).

ine strips (3 \times 5 mm) comprising the endometrium with the myometrium (ENDO/MYO) and myometrium (MYO) alone were resected, washed in saline and mounted between two stainless steel hooks in 5 ml of organ bath (Schuler Organ bath type 809; Hugo Sachs Electronic, Germany) under conditions of resting tension of 5 mN. The strips were kept in Krebs-Ringer solution of the following composition (mM/l): NaCl, 120.3; KCl, 5.9; CaCl₂, 2.5; MgCl₂, 1.2; NaHCO₃, 15.5; glucose, 11.5; 37°C, pH 7.4. The solution was maintained at 37°C and continuously saturated with a mixture 95% O₂ and 5% CO₂. The

recording was started after equilibration for at least 60 min. At the beginning of the experiment the strips were incubated with noradrenaline (Polfa, Poland) at dose of 10⁻⁶ M and acetylcholine (Sigma Aldrich, Germany) at a dose of 10⁻³ M to determine the viability of tissues and their usefulness to further study. Next, the effect of increasing concentrations of two analogues of LPA: oleoyl-sn-glycero-3-phosphate (L- α -LPA, a selective agonist of LPA₁ and LPA₂ receptors; 10⁻⁷ M; 10⁻⁶ M and 10⁻⁵ M; Sigma, USA) and 1-oleoyl-2-O-methyl-*rac*-glycerophosphothionate (OMPT, a selective agonist of LPA₃ receptor; 68 nM;

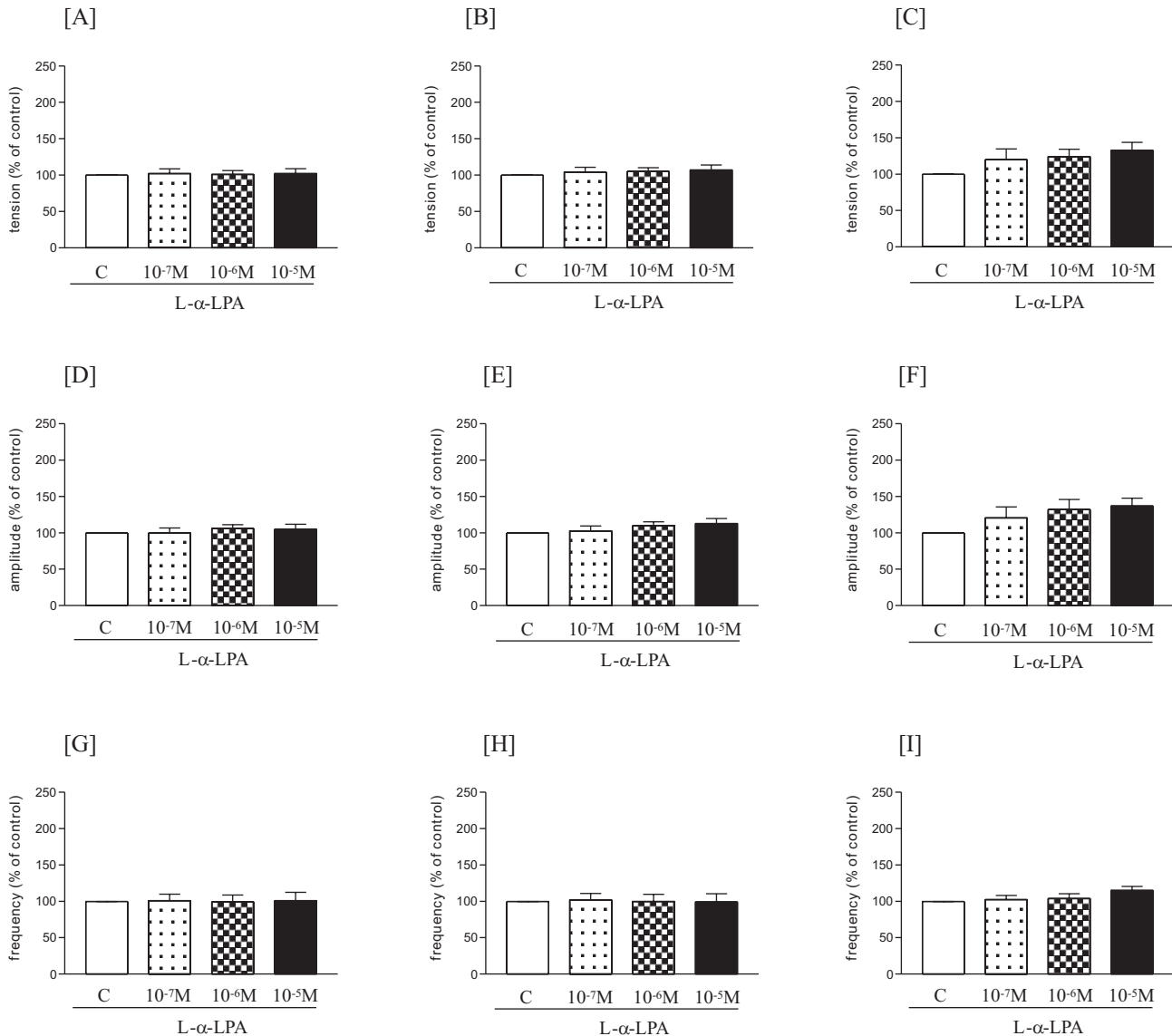


Fig. 2. Influence of oleoyl-sn-glycero-3-phosphate (L- α -LPA) on the tension ([A],[B],[C]) amplitude ([D],[E],[F]) and frequency ([G],[H],[I]) of the contraction of the porcine myometrium strips of the control group ([A],[D],[G]), uterine horn without embryos ([B],[E],[H]) and with the embryos ([C],[F],[I]). Uterine strips were collected on days 12-14 of the estrous cycle (control group) or pregnancy (groups with embryos and without embryos). Values (mean \pm SEM; $n = 5$ in each group) are expressed as a percentage of changes in the contractile activity before treatment (C).

136 nM and 680 nM; Cayman Chemical, USA) on the contractile activity was studied. The doses of the substances tested were determined on the basis of the preliminary study. Contractile activity was measured for 10 min after administration of each concentration of the substance examined and thereafter, tissue chambers were washed three times with 15 ml of phosphate buffer at 10 min intervals. In the end, to determine the viability of tissues NA and ACh were administered at doses given above. In the statistical analysis only those results were considered, for which the difference in the response to the stimulation by NA and ACh at the beginning and end of the study was smaller than 20%.

The measurements of the smooth muscle contraction were conducted using a force transducer (HSE F-30 type 372), and bridge coupler type 570, and the graphic recording was made using recorder (Hugo Sachs Elektronik) with HSE-ACADW software. All procedures involving animals were conducted in accordance with the Local Research Ethics Committee (67/2009 of 29.09.2009) national guidelines for agricultural animal care.

Statistical analysis

The numerical values of the contraction activity (tension, amplitude and frequency) of tissues before

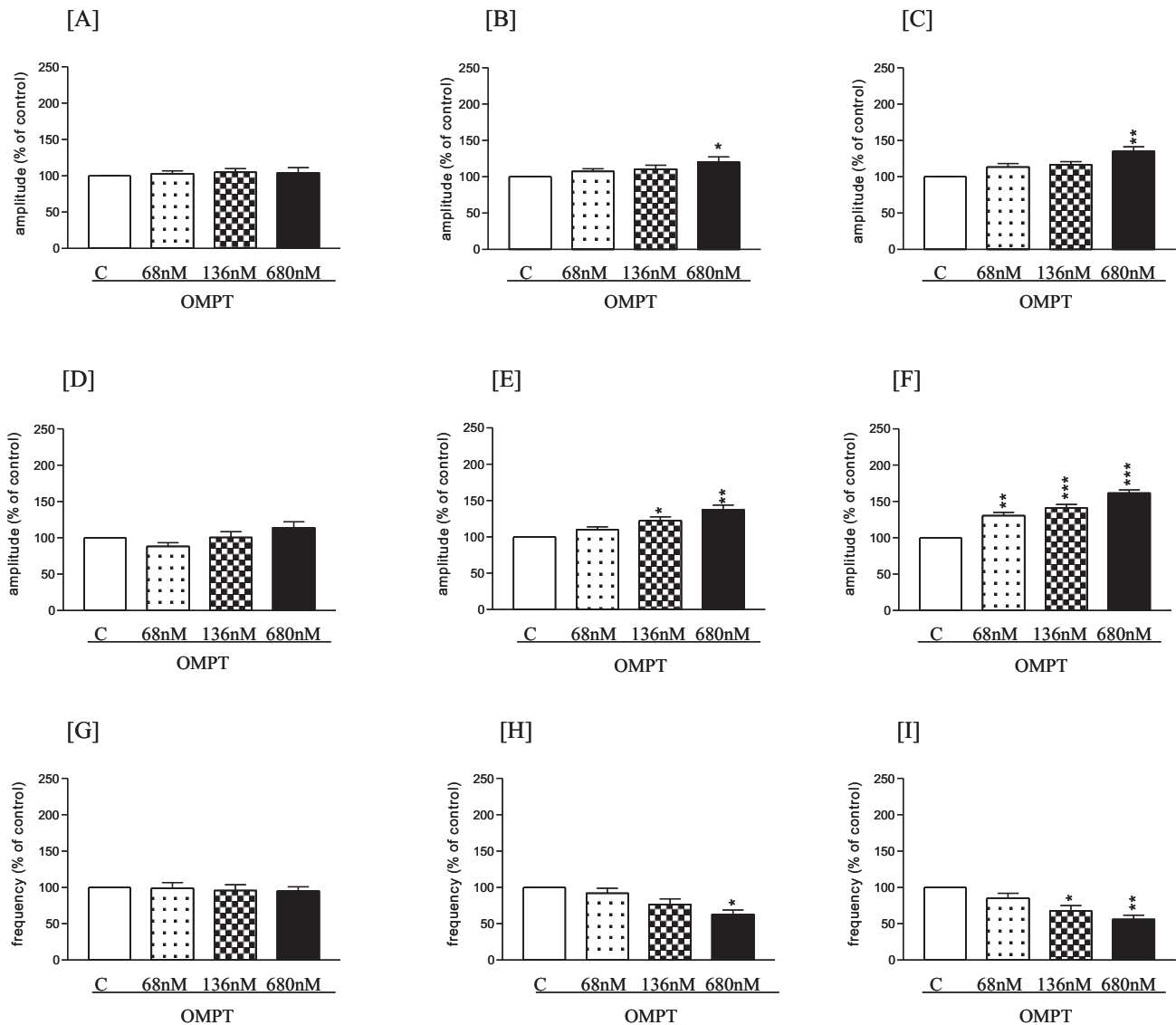


Fig. 3. Influence of 1-oleoyl-2-O-methyl-rac-glycerophosphothionate (OMPT) on the tension ([A],[B],[C]) amplitude ([D],[E],[F]) and frequency ([G],[H],[I]) of the contraction of the porcine endometrium/myometrium strips of the control group ([A],[D],[G]), uterine horn without embryos ([B],[E],[H]) and with the embryos ([C],[F],[I]). Uterine strips were collected on days 12-14 of the estrous cycle (control group) or pregnancy (groups with embryos and without embryos). Values (mean \pm SEM; n = 5 in each group) are expressed as a percentage of changes in the contractile activity before the treatment. * P < 0.05, ** P < 0.01, *** P < 0.001 as compared to the contractile activity before the treatment (C).

the application of biologically active substances were calculated for 7 min and accepted as 100%. The results calculated for 7-min period after treatments were expressed as a percentage (mean \pm SEM) of the contraction tension, amplitude and frequency before drug administration. The statistical significance of the differences obtained were assessed by one-way analysis of variance ANOVA (Graphpad PRISM 3.1; Graphpad Software, USA) followed by Bonferroni's Multiple Comparison Test. The differences at P < 0.05 were considered as statistically significant.

Results

Influence of L- α -LPA on the uterine contractile activity

In the ENDO/MYO uterine strips of the control group and uterine horn without the embryos, L- α -LPA at all examined doses did not cause significant changes (P > 0.05) in the contraction tension, amplitude and frequency (Figs. 1, 5B). In the ENDO/MYO strips of the uterine horn with the

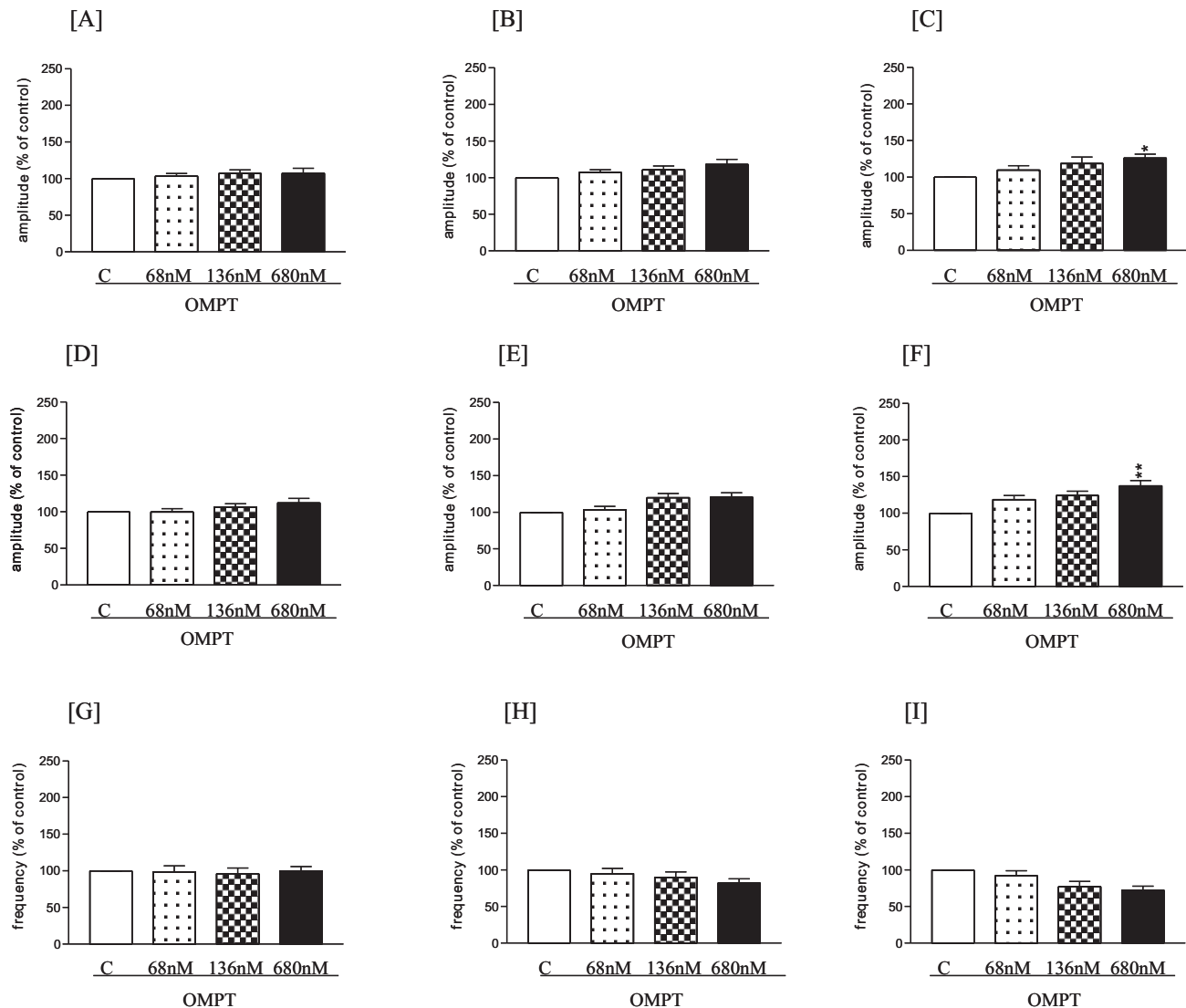


Fig. 4. Influence of 1-oleoyl-2-*O*-methyl-*rac*-glycerophosphothionate (OMPT) on the tension ([A],[B],[C]) amplitude ([D],[E],[F]) and frequency ([G],[H],[I]) of the contraction of the porcine myometrium strips of the control group ([A],[D],[G]), uterine horn without embryos ([B],[E],[H]) and with the embryos ([C],[F],[I]). Uterine strips were collected on days 12-14 of the estrous cycle (control group) or pregnancy (groups with embryos and without embryos). Values (mean \pm SEM; $n = 5$ in each group) are expressed as a percentage of changes in the contractile activity before the treatment. * $P < 0.05$, ** $P < 0.01$, as compared to the contractile activity before the treatment (C).

developing embryos, L- α -LPA at all doses increased ($P < 0.05$ -0.001) the tension (Fig. 1C), ($P < 0.01$, ($P < 0.001$) amplitude (Fig. 1F), while the frequency of contractions was enhanced only at the highest concentration of the studied agent (Figs. 1I, 5A).

In the MYO strips L- α -LPA did not affect significantly ($P > 0.05$) all parameters examined in both the control and experimental group (Fig. 2).

Influence of OMPT on the contractile activity of uteri

In the ENDO/MYO uterine strips of the control group, OMPT did not change significantly ($P > 0.05$)

all parameters examined (Fig. 3A,D,G). In the ENDO/MYO strips of the uterine horn without the developing embryos after OMPT administration, an increase in the contraction tension (at a dose of 680 nM; $P < 0.05$; Fig. 3B) and amplitude (at a doses of 136 and 680 nM; $P < 0.05$ and $P < 0.01$, respectively; Fig. 3E) was observed (Fig. 5D). However, the frequency of contractions after administration of the highest dose of OMPT was decreased ($P < 0.05$) (Fig. 1H). In the ENDO/MYO strips of the uterine horn with the embryos, OMPT increased ($P < 0.01$) the contraction tension at the highest dose (Fig. 1C), and amplitude at all doses examined (Fig. 1F), while the frequency of contractions at doses of 136 nM

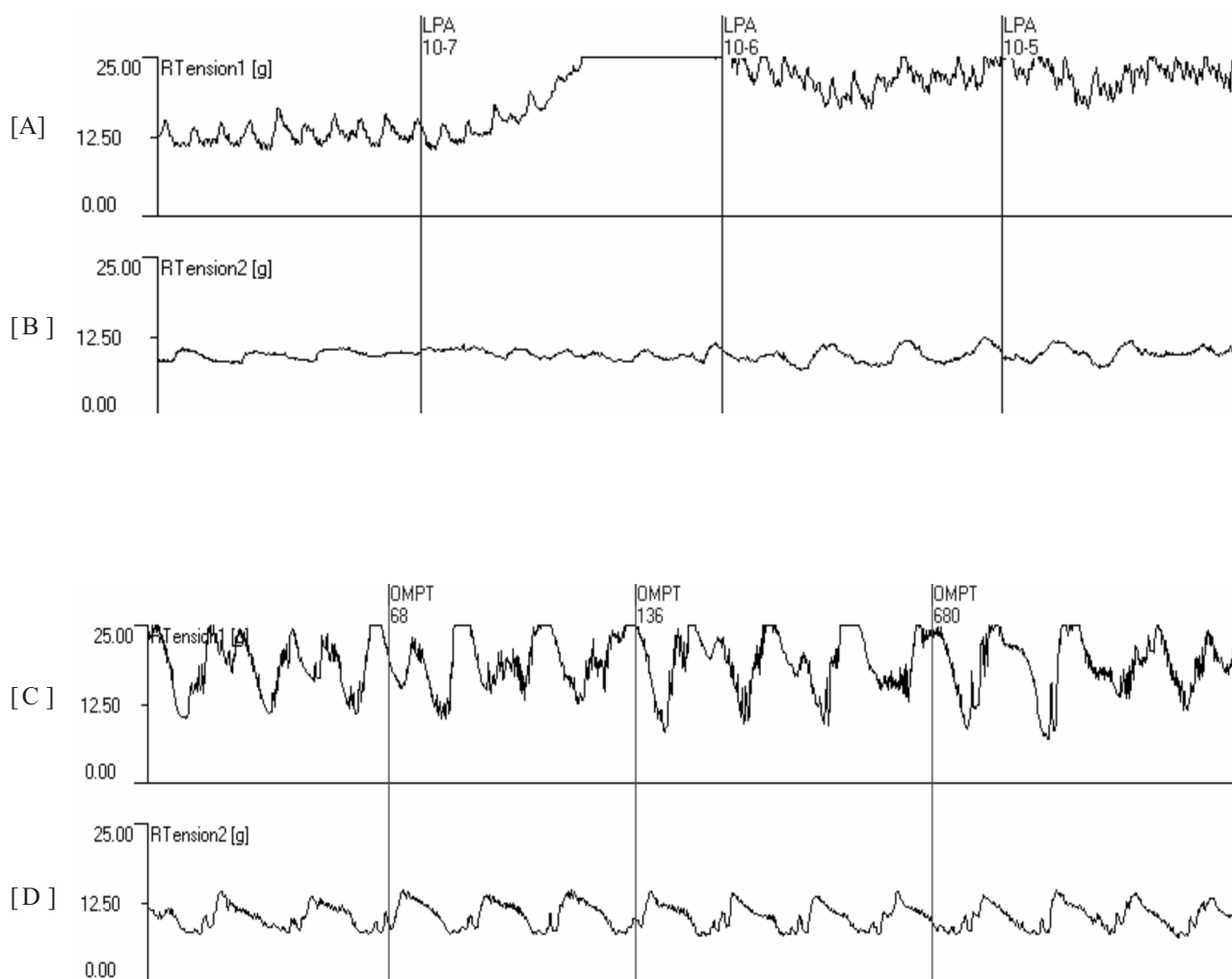


Fig. 5. Representative diagram showing the influence of oleoyl-sn-glycero-3-phosphate (L- α -LPA) and 1-oleoyl-2-*O*-methyl-*rac*-glycerophosphothionate (OMPT) on the contractile activity of the porcine endometrium/myometrium strips collected on days 12-14 of pregnancy. Each panel shows the contractile activity in the response to increasing doses of L- α -LPA (10^{-7} – 10^{-5} M) in the uterine horn with the embryos (Fig. 5A) and without embryos (Fig. 5B) and OMPT (68 nM; 136 nM and 680 nM) in the uterine horn with the embryos (Fig. 5C) and without embryos (Fig. 5D).

($P < 0.05$) and 680 nM ($P < 0.01$) (Fig. 3I) was decreased (Fig. 5C). In the MYO strips of the uterine horn with the developing embryos, OMPT administration at the highest dose caused an increase in the contraction tension ($P < 0.05$; Fig. 4C) and amplitude ($P < 0.01$; Fig. 4F) while the frequency of contractions was not affected.

Discussion

Since the first report that LPA induces cell proliferation and differentiation similar to that induced by growth factors via G protein-coupled receptors (Van Corven et al. 1989), many studies have shown that LPA participates in diverse physiological functions

such as angiogenesis, neuronal and cardiovascular development, immunomodulation, smooth muscle contraction and relaxation, and tumorigenesis (Ishii et al. 2004). There is not much data about LPA function in the uterus but recent reports have suggested that LPA receptor signaling plays an important role in the embryo implantation process (Ye et al. 2005). In the studies conducted on mice (Ye et al. 2005, Hama et al. 2007) it was found that improper functioning of LPA in the uterus may result in delayed implantation, improper embryo spacing along the uterine horns and disturbed embryo development. In the present study we showed that L- α -LPA increased the contractile activity in the ENDO/MYO from the uterine horn with the developing embryos but did not affect the contractility in the uterine horn without the embryos. OMPT

increased the contractile activity in ENDO/MYO strips of both uterine horns with and without the embryos, and in MYO strips of the uterine horn with the embryos. It may suggest that activation of LPA₁ and LPA₂ is possible only in the presence of embryos, while OMPT acts rather through LPA₃ with the stronger effect in the uterus of pregnant pigs (regardless of the embryo presence in the uterine horn) comparing to the cyclic uterus. Our results are consistent with the previous study performed by Hama et al. (2007). They found that that selective agonist of LPA₃ (1-oleoyl LPA) caused contraction of smooth muscles of the pregnant mouse uterus. However, such effect was not observed in the uterus of mice lacking gene responsible for LPA₃ synthesis but with the preserved contraction activity confirmed by reaction to acetylcholine. Moreover, Kaminska et al. (2008) showed higher level of mRNA for LPA₃ gene in the porcine endometrial tissue on days 6-7 of pregnancy as compared to the cyclic uterus. Higher mRNA level was also found in the uterine horn containing embryos compared to the contralateral horn, where embryos did not develop. These results imply the important role of receptor LPA₃ during early pregnancy.

In conclusion, direct activation of LPA₁, LPA₂ i LPA₃ in the porcine myometrium during early pregnancy results in the enhanced contraction activity of smooth muscles. Furthermore, it was found that not only LPA₃ but also two other isoforms of the receptor, LPA₁ and LPA₂, participate in contractions of the uterus in pregnant pigs. The present data indicate the important role of LPA and its receptors in the induction of uterine contractions what is the main factor influencing embryo spacing.

References

- An S, Bleu T, Hallmark OG, Goetzl EJ (1998) Characterization of a novel subtype of human G protein-coupled receptor for lysophosphatidic acid. *J Biol Chem* 273: 7906-7910.
- Bandoh K, Aoki J, Hosono H, Kobayashi S, Kobayashi T, Murakami-Murofushi K, Tsujimoto M, Arai H, Inoue K (1999) Molecular cloning and characterization of a novel human G-protein-coupled receptor, EDG7, for lysophosphatidic acid. *J Biol Chem* 274: 27776-27785.
- Budnik LT, Mukhopadhyay AK (2002) Lysophosphatidic acid and its role in reproduction. *Biol Reprod* 66: 859-865.
- Cao J, Shayibuzhati M, Tajima T, Kitazawa T, Taneike T (2002) In vitro pharmacological characterization of the prostanoid receptor population in the non-pregnant porcine myometrium. *Eur J Pharmacol* 442: 115-123.
- Carnahan KG, Prince BC, Ludwig TE, Uzumcu M, Ewans MA, Miranda MA (1999) Effect of oxytocin on concentration of PGF_{2α} in the uterine lumen and subsequent endometrial responsiveness to oxytocin in pigs. *J Reprod Fertil* 117: 207-212.
- Contos JJ, Fukushima N, Weiner JA, Kaushal D, Chun J (2000a) Requirement for the lpa1 lysophosphatidic acid receptor gene in normal suckling behavior. *Proc Natl Acad Sci USA* 97: 13384-13389.
- Contos JJ, Ishii I, Chun J (2000b) Lysophosphatidic acid receptors. *Mol Pharmacol* 58: 1188-1196.
- Contos JJ, Ishii I, Fukushima N, Kingsbury MA, Ye X, Kawamura S, Brown JH, Chun J (2002) Characterization of lpa(2) (Edg4) and lpa(1)/lpa(2) (Edg2/Edg4) lysophosphatidic acid receptor knockout mice: signaling deficits without obvious phenotypic abnormality attributable to lpa(2). *Mol Cell Biol* 22: 6921-6929.
- Franczak A, Bogacki M (2009) Local and systemic effects of embryos on uterine tissues during early pregnancy in pigs. *J Reprod Dev* 55: 262-272.
- Gardell SE, Dubin AE, Chun J (2006) Emerging medicinal roles for lysophospholipid signaling. *Trends Mol Med* 12: 65-75.
- Hama K, Aoki J, Inoue A, Endo T, Amano T, Motoki R, Kanai M, Ye X, Chun J, Matsuki N, Suzuki H, Shibasaki M, Arai H (2007) Embryo spacing and implantation timing are differentially regulated by LPA₃-mediated lysophosphatidic acid signaling in mice. *Biol Reprod* 77: 954-959.
- Inoue M, Rashid MH, Fujita R, Contos JJ, Chun J, Ueda H (2004) Initiation of neuropathic pain requires lysophosphatidic acid receptor signaling. *Nat Med* 10: 712-718.
- Isaka M, Takaoka K, Yamada Y, Abe Y, Kitazawa T, Taneike T (2000) Characterization of functional endothelin receptors in the porcine myometrium. *Peptides* 21: 543-551.
- Ishii I, Fukushima N, Ye X, Chun J (2004) Lysophospholipid receptors: signaling and biology. *Annu Rev Biochem* 73: 321-354.
- Jana B, Jaroszewski JJ, Kucharski J, Koszykowska M, Górka J, Markiewicz W (2010) Participation of prostaglandin E2 in contractile activity of inflamed porcine uterus. *Acta Vet Brno* 79: 249-259.
- Kaleczyc J (1994) Effect of estradiol and progesterone on noradrenaline content in nerves of the oviduct, uterus and vagina in ovariectomized pigs. *Folia Histochem Cytobiol* 32: 119-126.
- Kamińska K, Wasielek M, Bogacka I, Blitek M, Bogacki M (2008) Quantitative expression of lysophosphatidic acid receptor 3 gene in porcine endometrium during the periimplantation period and estrous cycle. *Prostaglandins Other Lipid Mediat* 85: 26-32.
- Kitazawa T, Shishido H, Sato T, Taneike T (1997) Histamine mediates the muscle layer-specific responses in the isolated swine myometrium. *J Vet Pharmacol Ther* 20: 187-197.
- Kitazawa T, Nakagoshi K, Teraoka H, Taneike T (2001a) 5-HT(7) receptor and β₂-adrenoceptor share in the inhibition of porcine uterine contractility in a muscle layer-dependent manner. *Eur J Pharmacol* 433: 187-197.
- Kitazawa T, Kajiwara T, Kiuchi A, Hatakeyama H, Taneike T (2001b) Muscle layer- and region-dependent distributions of oxytocin receptors in the porcine myometrium. *Peptides* 22: 963-974.
- Kitazawa T, Hatakeyama H, Cao J, Taneike T (2003) Pregnancy-associated changes in responsiveness of the por-

- cine myometrium to bioactive substances. *Eur J Pharmacol* 469: 135-144.
- Kucharski J, Jaroszewski J, Jana B, Gorska J, Kozłowska A, Markiewicz W (2007) The influence of inflammatory process on prostaglandin F_{2α} contractile activity in porcine uterus. *J Anim Feed Sci* 16: 654-667.
- Kurowicka B, Franczak A, Oponowicz A, Kotwica G (2005) In vitro contractile activity of porcine myometrium during luteolysis and early pregnancy: effect of oxytocin and progesterone. *Reprod Biol* 5: 151-169.
- Langendijk P, Bouwman EG, Soede NM, Taverne MA, Kemp B (2002) Myometrial activity around estrus in sows: spontaneous activity and effects of estrogens, cloprostenol, seminal plasma and clenbuterol. *Theriogenology* 57: 1563-1577.
- Langendijk P, Soede NM, Kemp B (2005) Uterine activity, sperm transport, and the role of boar stimuli around insemination in sows. *Theriogenology* 63: 500-513.
- Li C, Dandridge KS, Di A, Marrs KL, Harris EL, Roy K, Jackson JS, Makarova NV, Fujiwara Y, Farrar PL, Nelson DJ, Tigyi GJ, Naren AP (2005) Lysophosphatidic acid inhibits cholera toxin-induced secretory diarrhea through CFTR-dependent protein interactions. *J Exp Med* 202: 975-986.
- Liliom K, Bittman R, Swords B, Tigyi G (1996b) N-Palmitoyl-serine and N-palmitoyl-tyrosine phosphoric acids are selective competitive antagonists of the lysophosphatidic acid receptors. *Mol Pharmacol* 50: 616-623.
- Liliom K, Murakami-Murofushi K, Kobayashi S, Murofushi H, Tigyi G (1996a) *Xenopus* oocytes express multiple receptors for LPA-like lipid mediators. *Am J Physiol* 270: C772-777.
- Łakomy M (1987) Adrenergic and cholinergic innervation of female reproductive organs in pig in particular periods of ovarian, pregnancy and lactation cycles. *Zeszyty Problenowe PNR* 339: 89-104.
- Majewski M, Sienkiewicz W, Kaleczyc J, Mayer B, Czaja K, Łakomy M (1995) The distribution and co-localization of immunoreactivity to nitric oxide synthase, vasoactive intestinal polypeptide and substance P within nerve fibres supplying bovine and porcine female genital organs. *Cell Tissue Res* 281: 445-464.
- Markiewicz W, Jaroszewski JJ, Bossowska A, Majewski M (2003) NPY: its occurrence and relevance in the female reproductive system. *Folia Histochem Cytobiol* 41: 183-192.
- Moolenaar WH (1995) Lysophosphatidic acid, a multifunctional phospholipid messenger. *J Biol Chem* 270: 12949-12952.
- Noguchi K, Ishii S, Shimizu T (2003) Identification of p2y₉/GPR23 as a novel G protein-coupled receptor for lysophosphatidic acid, structurally distant from the Edg family. *J Biol Chem* 278: 25600-25606.
- Seo H, Kim M, Choi Y, Lee CK, Ka H (2008) Analysis of lysophosphatidic acid (LPA) receptor and LPA-induced endometrial prostaglandin-endoperoxide synthase 2 expression in the porcine uterus. *Endocrinology* 149: 6166-6175.
- Sukjumlong S, Persson E, Dalin AM, Janson V, Sahlin L (2009) Messenger RNA levels of estrogen receptors α and β and progesterone receptors in the cyclic and inseminated/early pregnant sow uterus. *Anim Reprod Sci* 112: 215-228.
- Taneike T, Miyazaki H, Nakamura H, Ohga A (1991) Autonomic innervation of the circular and longitudinal layers in swine myometrium. *Biol Reprod* 45: 831-840.
- Thilander G, Rodriguez-Martinez H (1989) Distribution of adrenergic and cholinergic nerves in the porcine myometrium during the oestrous cycle. A histochemical investigation. *Zentralbl Veterinarmed A* 36: 276-284.
- Tigyi G, Dyer DL, Miledi R (1994) Lysophosphatidic acid possesses dual action in cell proliferation. *Proc Natl Acad Sci USA* 91: 1908-1912.
- Tigyi G, Miledi R (1992) Lysophosphatidates bound to serum albumin activate membrane currents in *Xenopus* oocytes and neurite retraction in PC12 pheochromocytoma cells. *J Biol Chem* 267: 21360-21367.
- Tokumura A, Fujimoto H, Yoshimoto O, Nishioka Y, Miyake M, Fukuzawa K (1999) Production of lysophosphatidic acid by lysophospholipase D in incubated plasma of spontaneously hypertensive rats and Wistar Kyoto rats. *Life Sci* 65: 245-253.
- Tokumura A, Harada K, Fukuzawa K, Tsukatani H (1986) Involvement of lysophospholipase D in the production of lysophosphatidic acid in rat plasma. *Biochim Biophys Acta* 875: 31-38.
- Van Corven EJ, Groenink A, Jalink K, Eichholtz T, Moolenaar WH (1989) Lysophosphatidate-induced cell proliferation: identification and dissection of signaling pathways mediated by G proteins. *Cell* 59: 45-54.
- Wasielak M, Glowacz M, Kaminska K, Waclawik A, Bogacki M (2008) The influence of embryo presence on prostaglandins synthesis and prostaglandin E₂ and F_{2α} content in corpora lutea during periimplantation period in the pig. *Mol Reprod Dev* 75: 1208-1216.
- Xie Y, Meier KE (2004) Lysophospholipase D and its role in LPA production. *Cell Signal* 16: 975-981.
- Ye X, Hama K, Contos JJ, Anliker B, Inoue A, Skinner MK, Suzuki H, Amano T, Kennedy G, Arai H, Aoki J, Chun J (2005) LPA₃-mediated lysophosphatidic acid signalling in embryo implantation and spacing. *Nature* 435: 104-108.